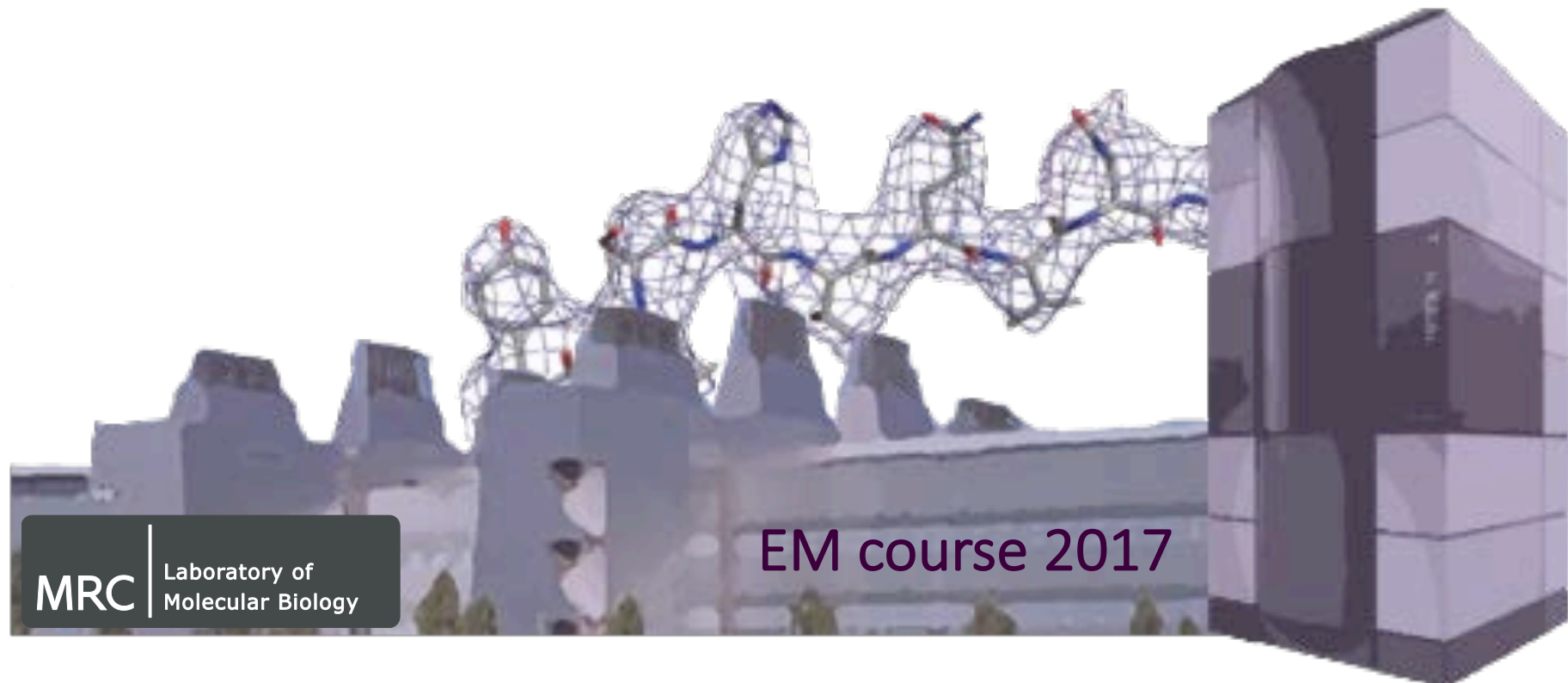


# Tomography and Subtomogram Averaging

John Briggs



**MRC** | Laboratory of  
Molecular Biology

EM course 2017

# What do we need to get a 3D structure?

Sample preparation methods

A transmission electron microscope

Different views of our object of interest

Computational approaches for producing a 3D reconstruction from 2D projections

Methods for validation and interpretation of the 3D structure

# What do we need to get a 3D structure?

Sample preparation methods

A transmission electron microscope

Different views of our object of interest

Computational approaches for producing a 3D reconstruction from 2D projections

Methods for validation and interpretation of the 3D structure

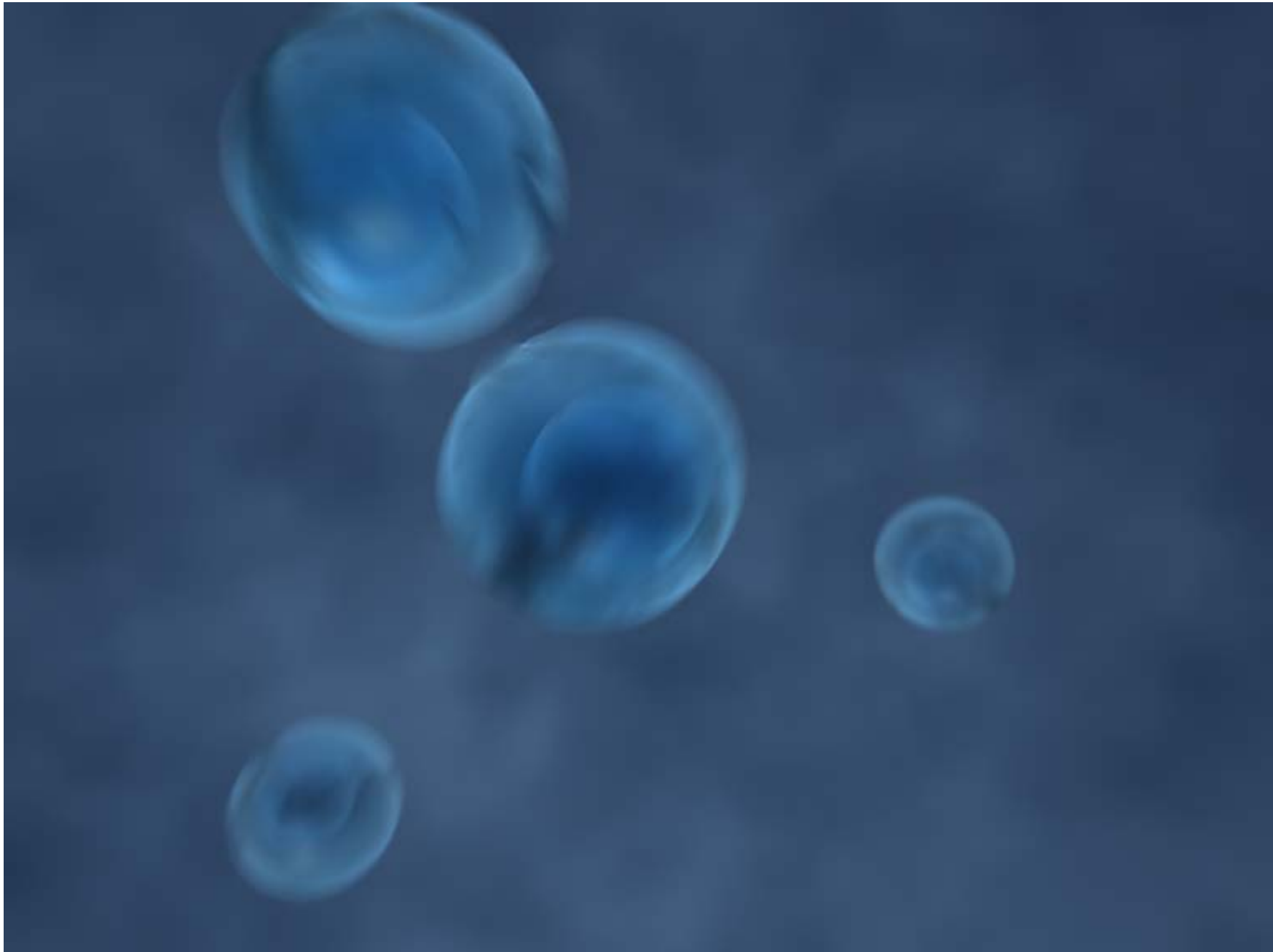
We need different views of our object



# Tomography?

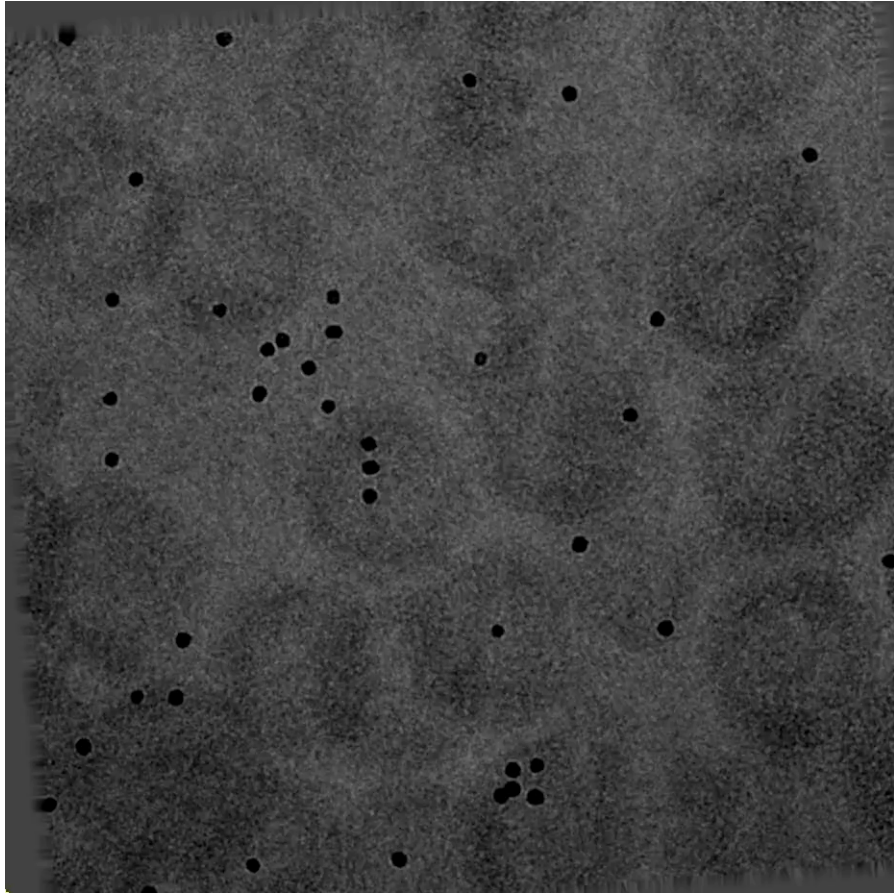
In single-particle methods we obtain different views of our object of interest by imaging many different copies that are oriented differently relative to the electron beam.

In tomography we obtain different views by physically rotating the sample in the microscope

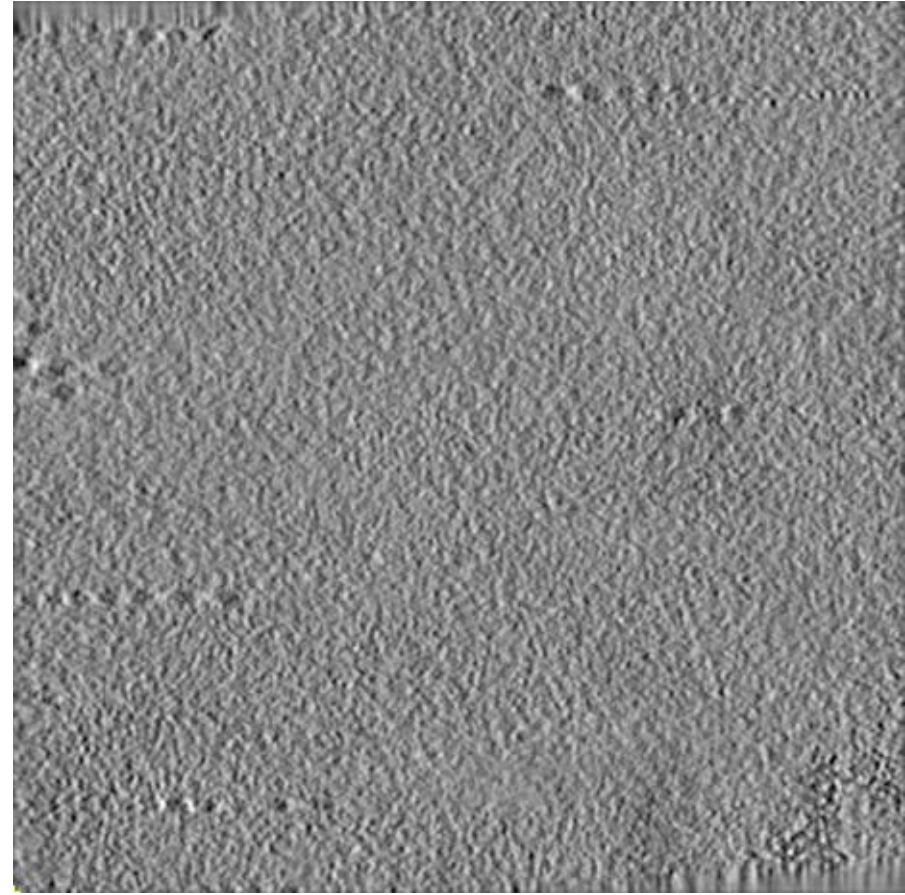


Movie from the Baumeister lab, Max-Planck Institute for Biochemistry

# Tilt series and tomogram



Tilt-series



Tomogram

# Why do tomography?

Because our sample is a unique structure (bits of cells, tissues, viruses etc.)

Because our sample is within a complex environment



# Topics to be covered

Sample preparation methods

Data collection and microscope requirements

Alignment and reconstruction

Subtomogram averaging

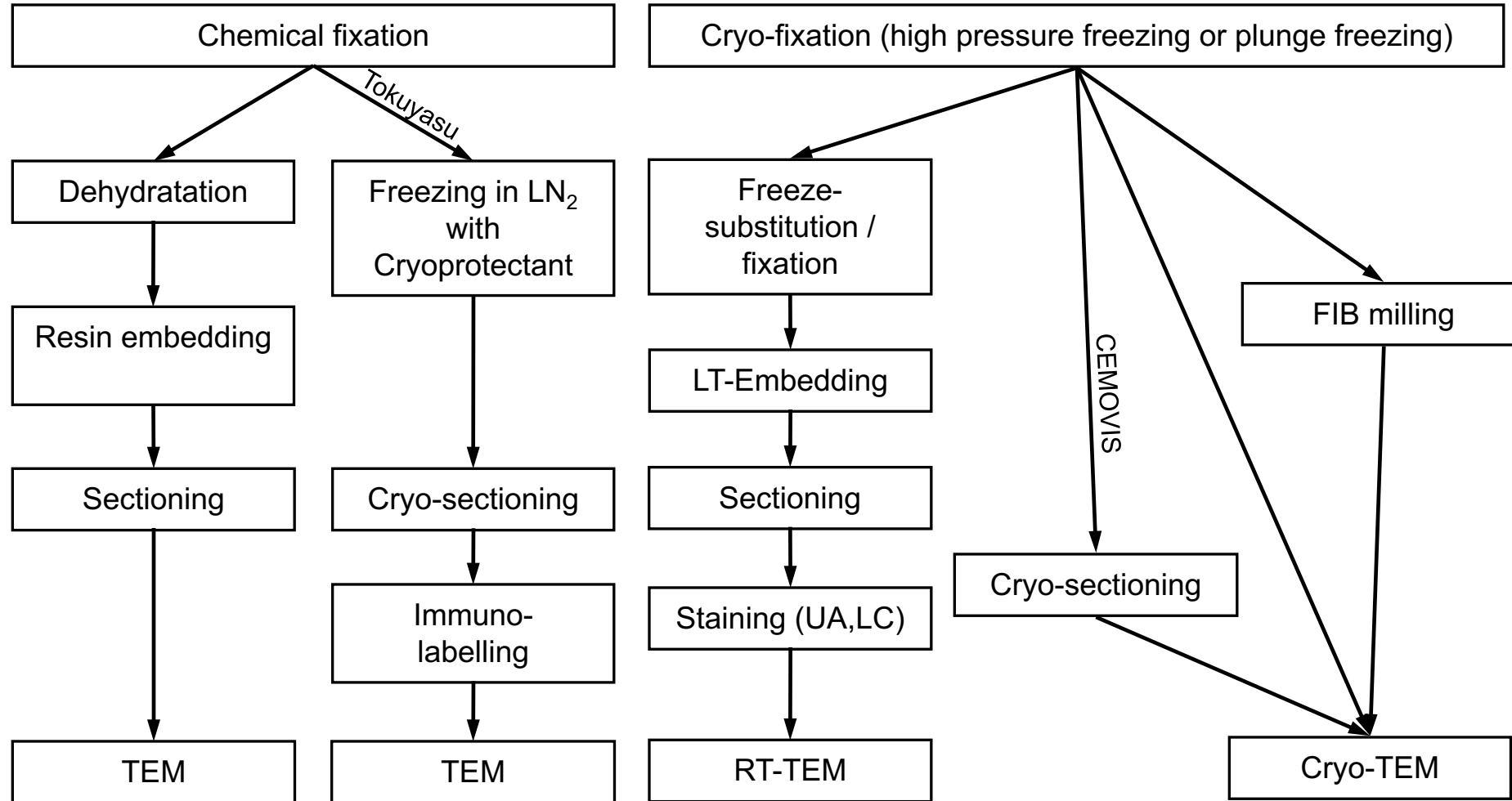
# Sample preparation

Sample must have structure preserved

Sample must be thin

Sample must be stable in vacuum

# Cellular sample preparation methods



# Cryo-sectioning (CEMOVIS)



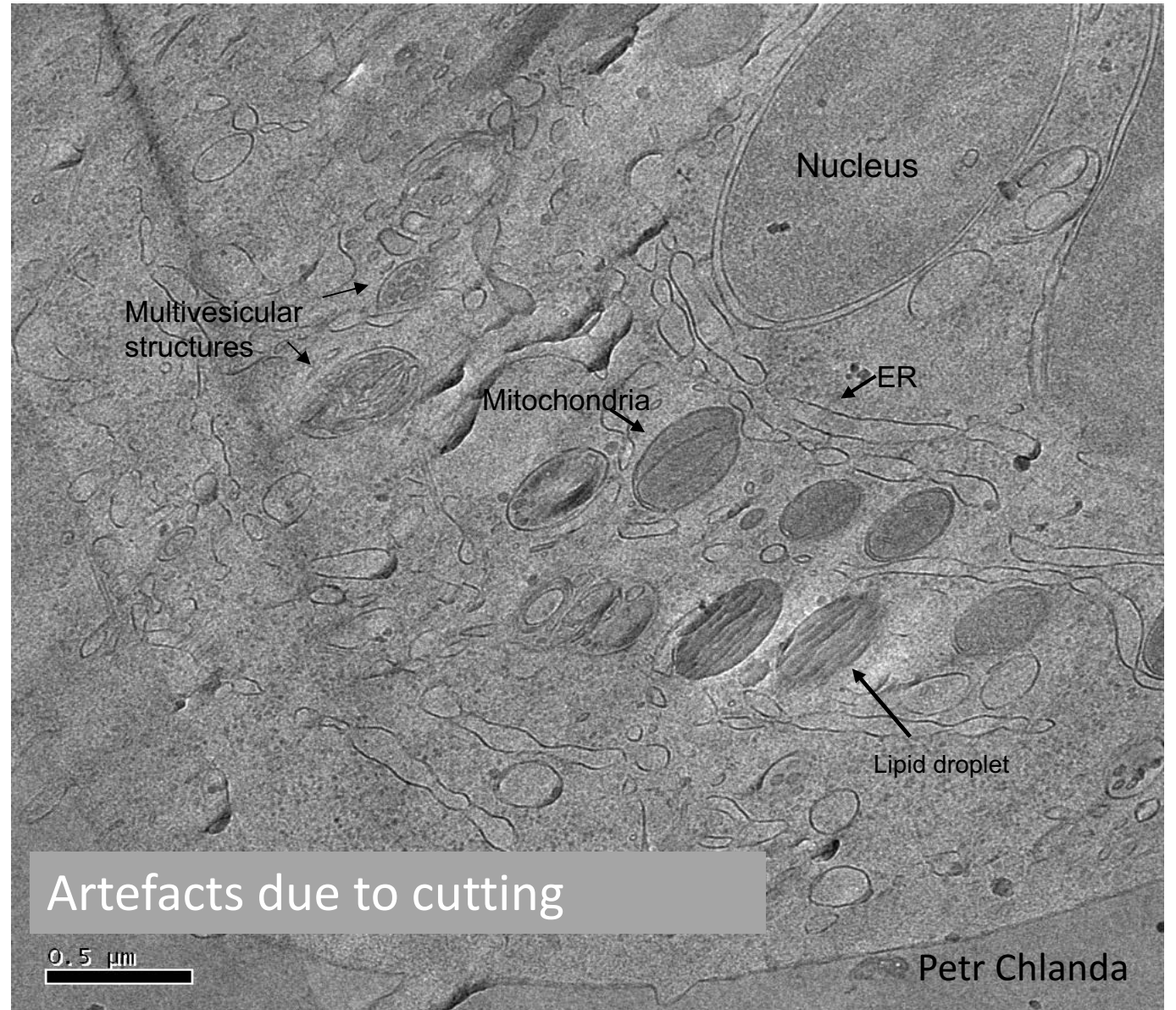
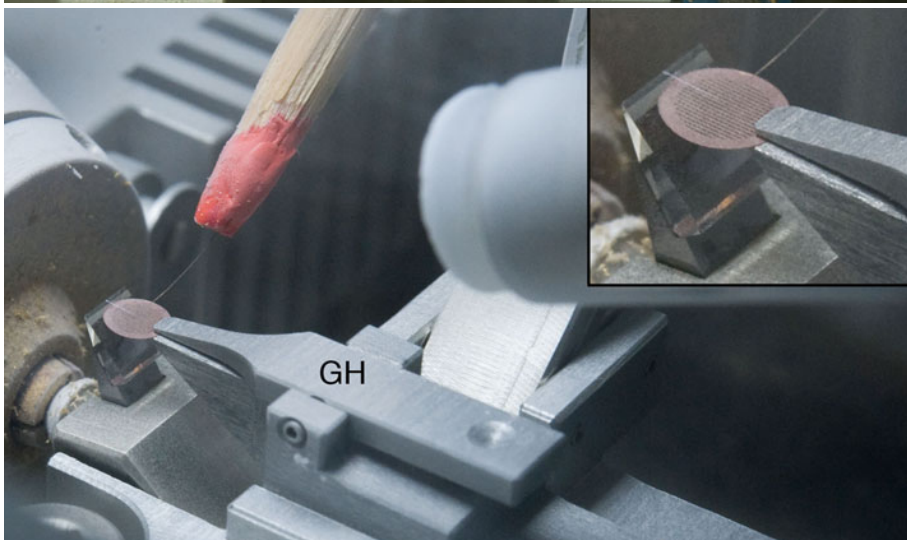
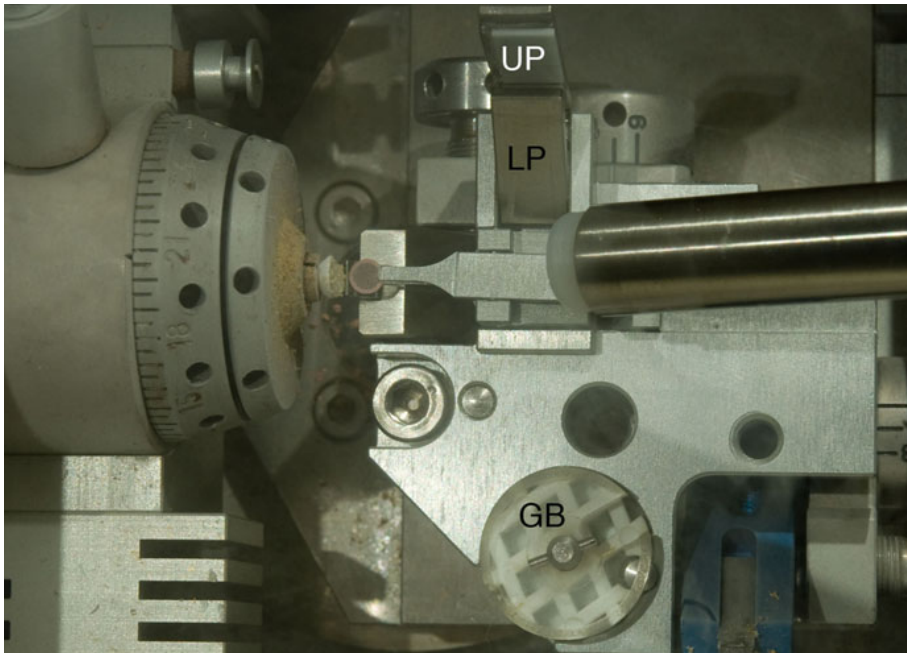
High-pressure freezer



Cryo-microtome

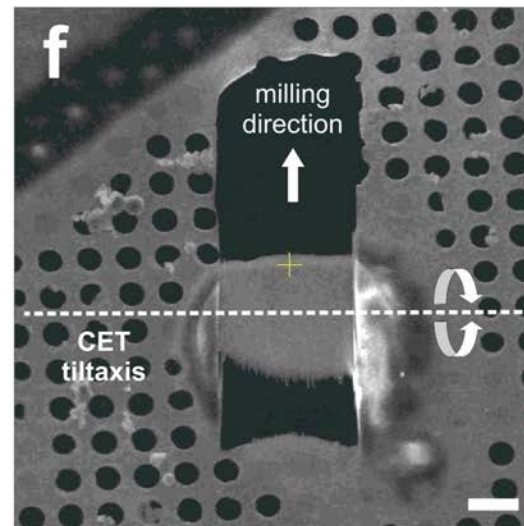
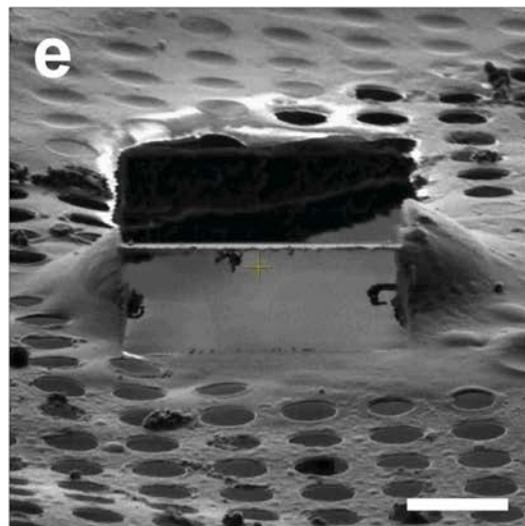
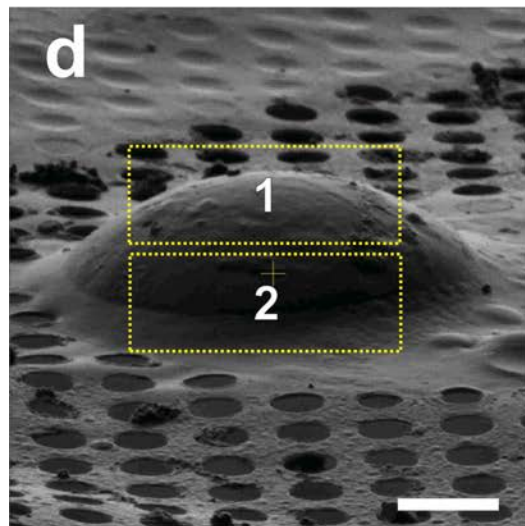
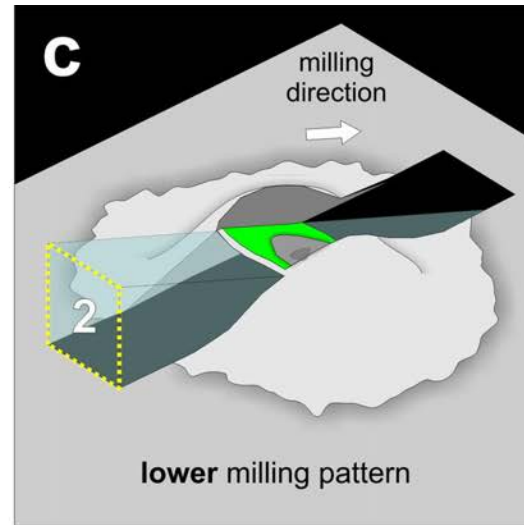
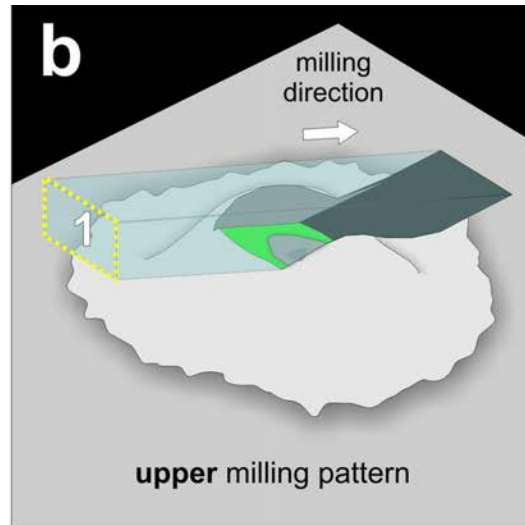
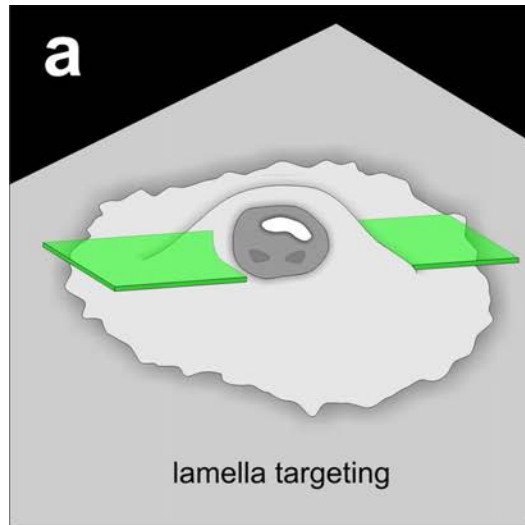


# Cryo-sectioning (CEMOVIS)



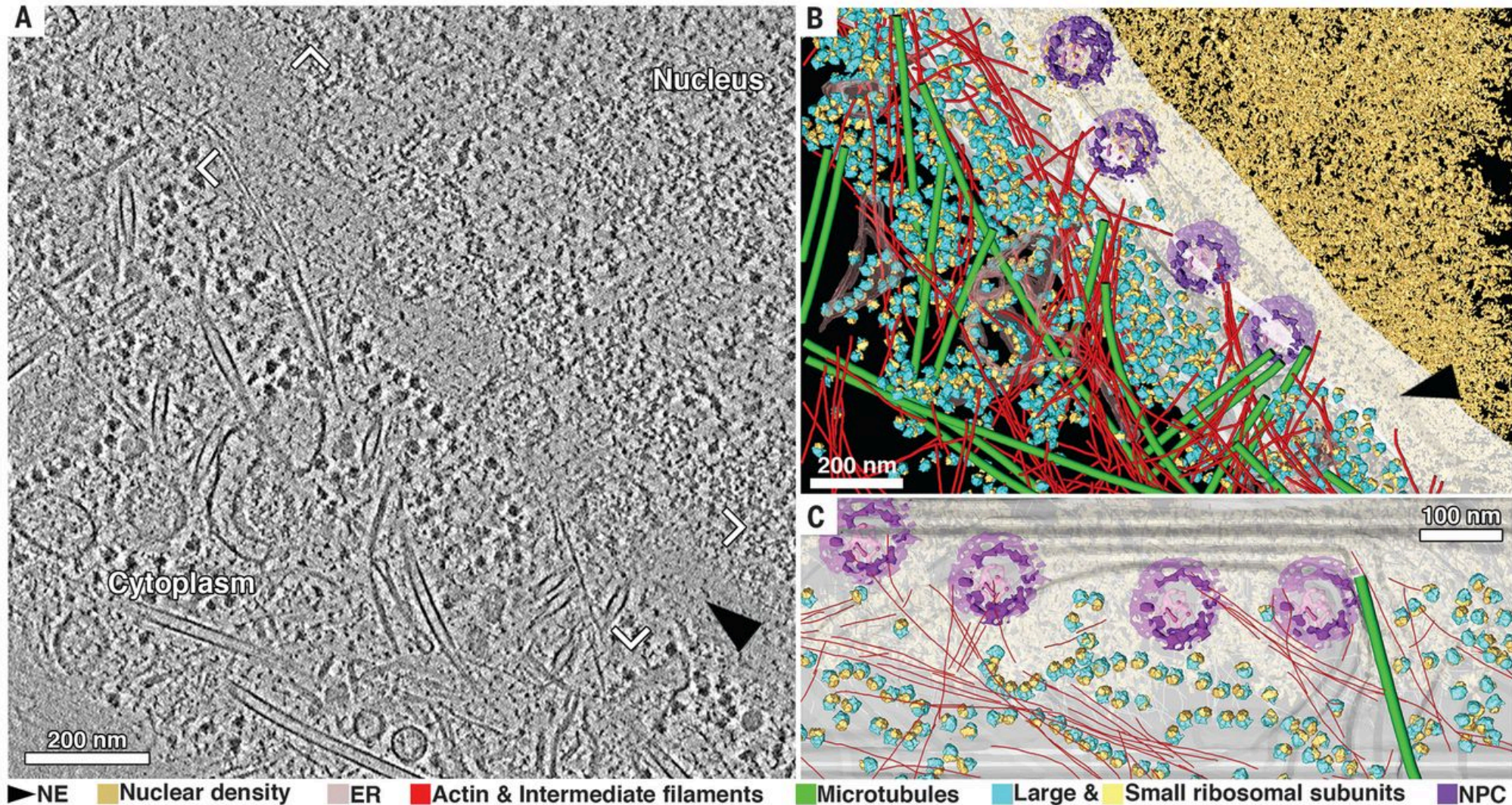


# Sample preparation - FIBSEM



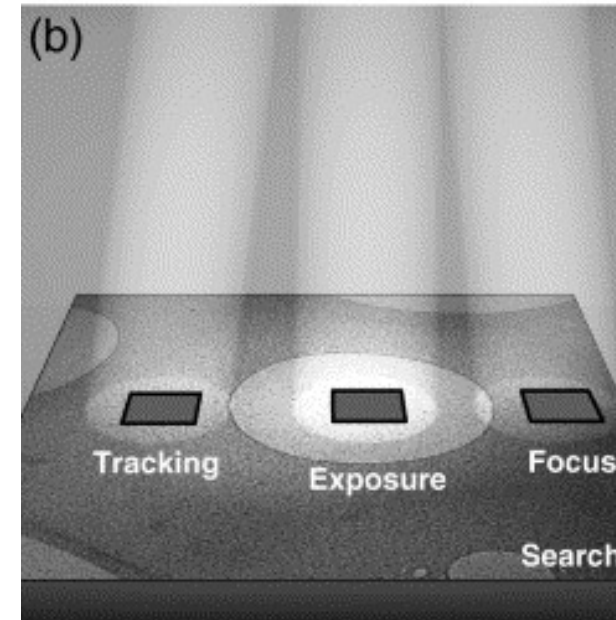
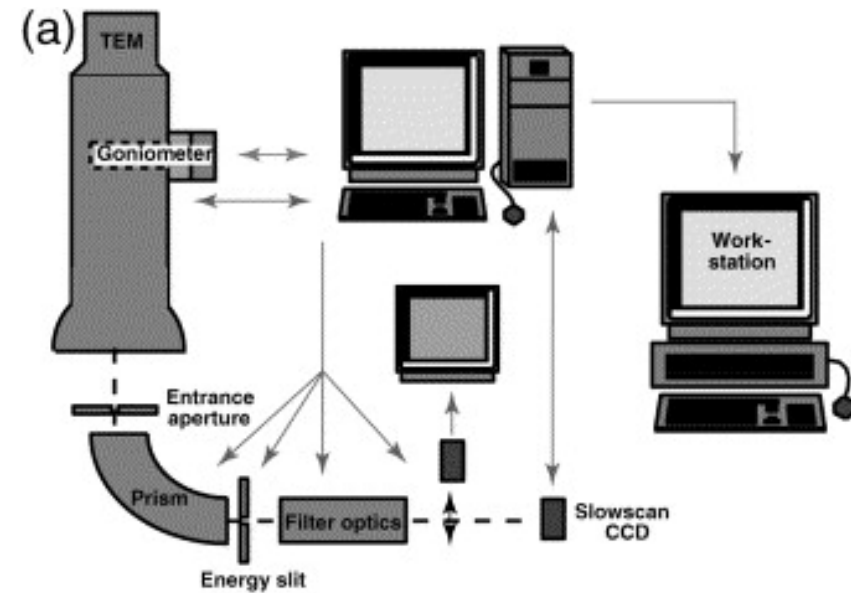
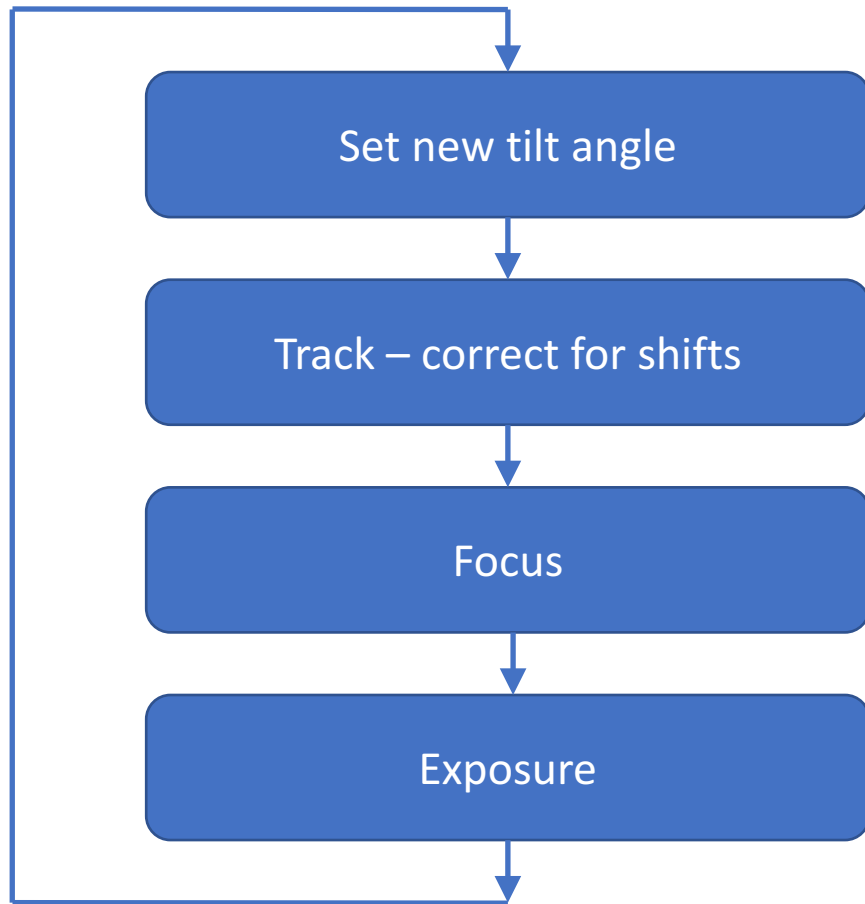


# Sample preparation - FIBSEM





# Data collection





# Microscope requirements

(for cryo-electron tomography of thicker samples)

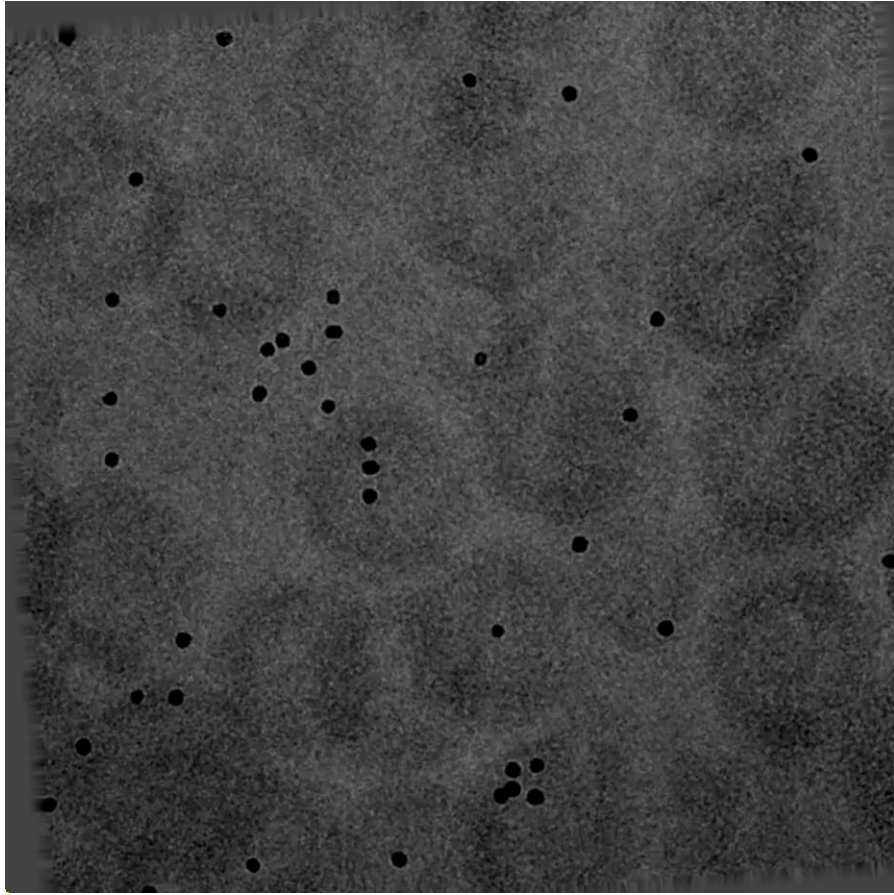
High-voltage (ideally 300kV)

Energy filter

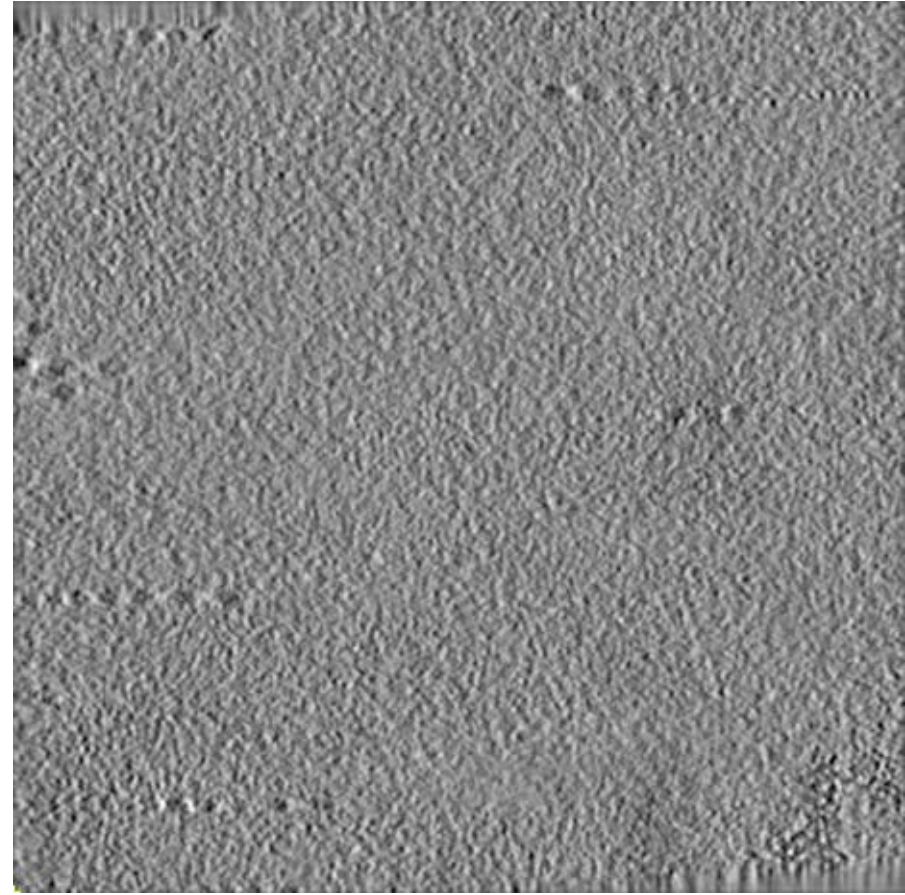
Very stable and well-calibrated microscope stage

Appropriate software

# Tilt series and tomogram



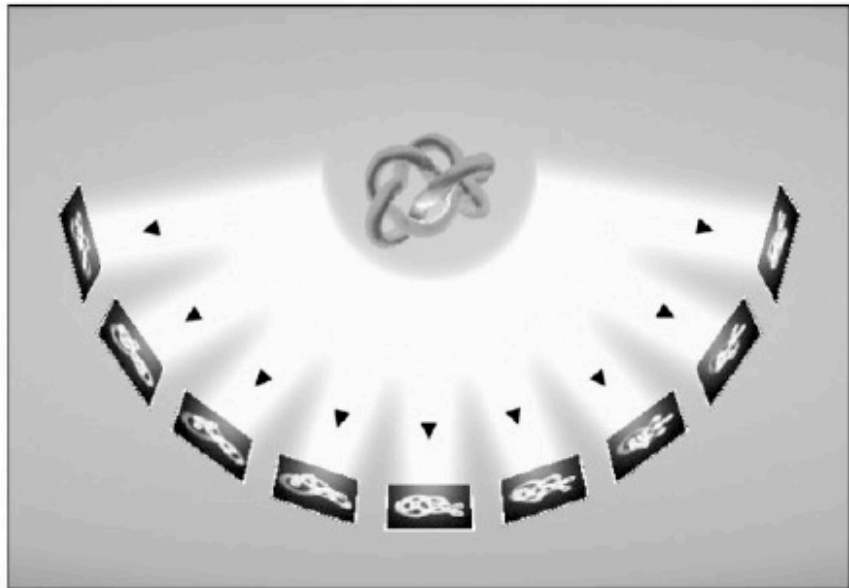
Tilt-series



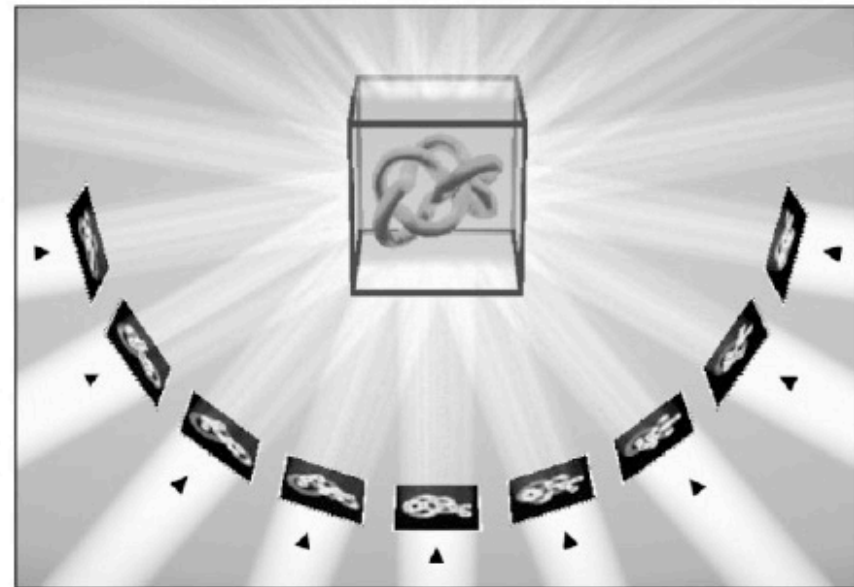
Tomogram

# Forward and back projection

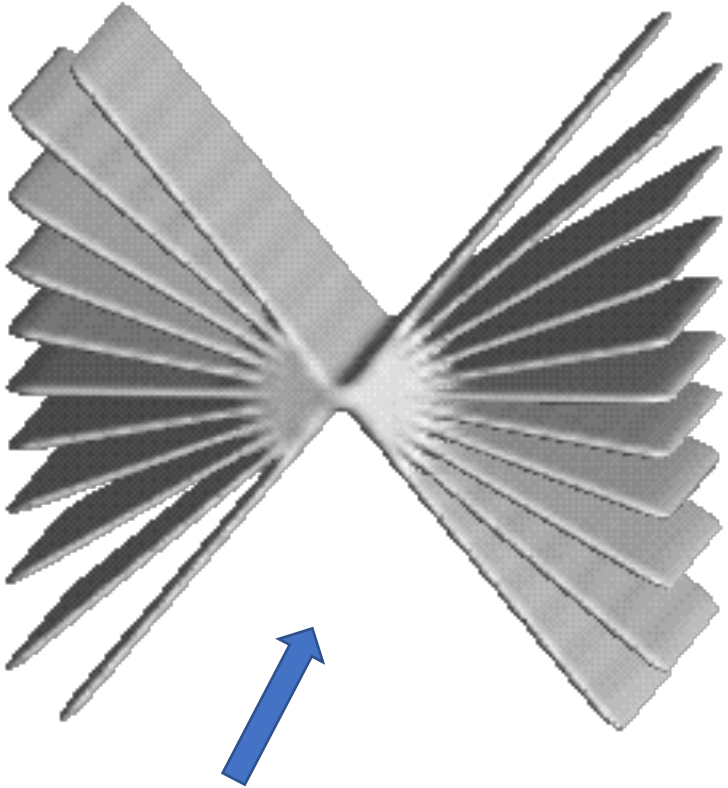
3D object -> 2D projections



2D projections -> 3D reconstruction

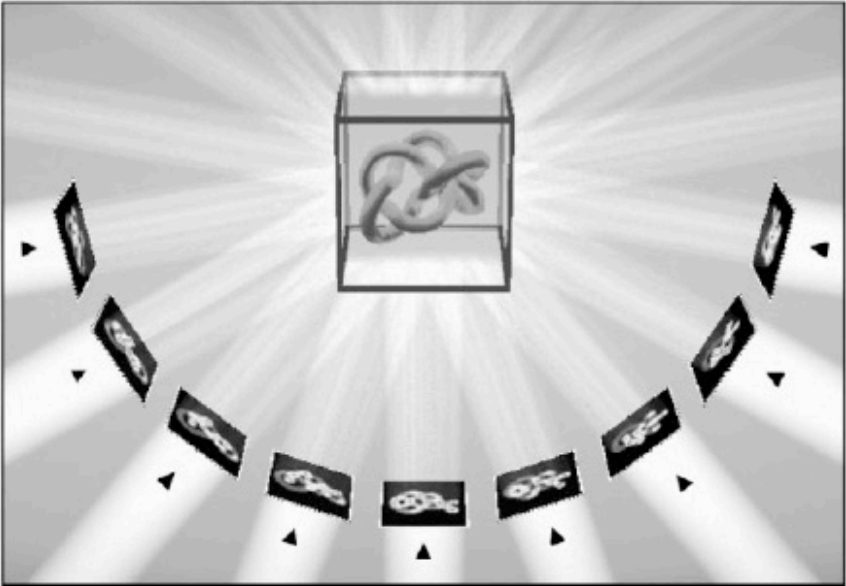


# The data is incomplete: the missing wedge

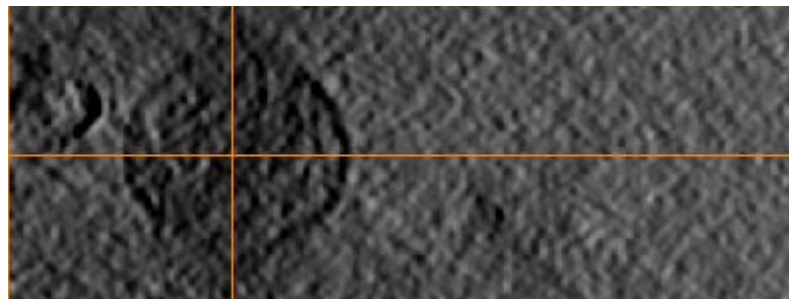
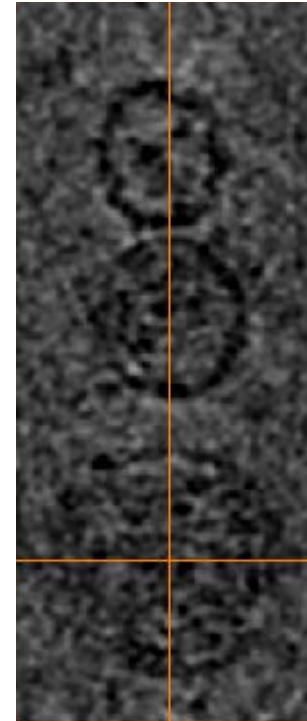
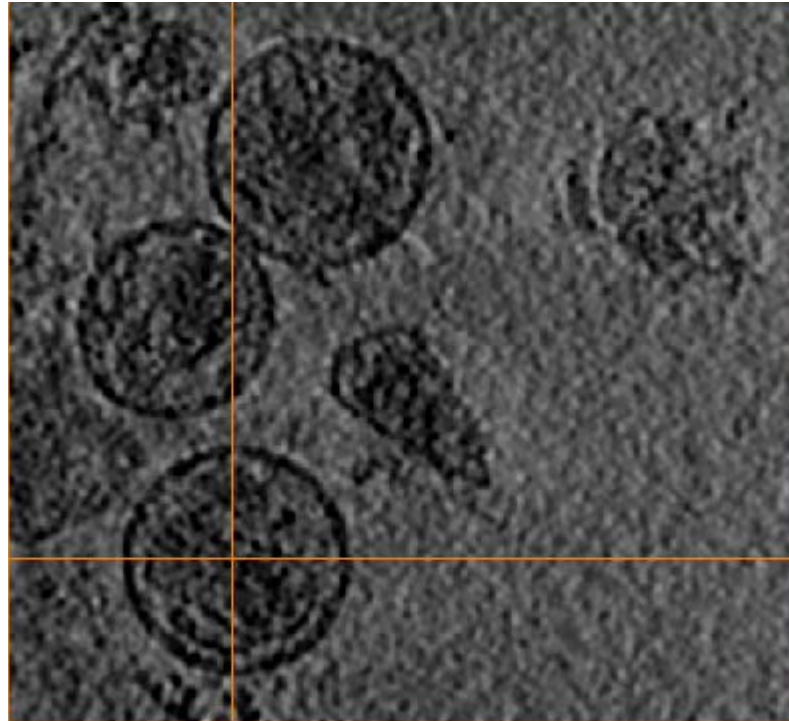


Missing information

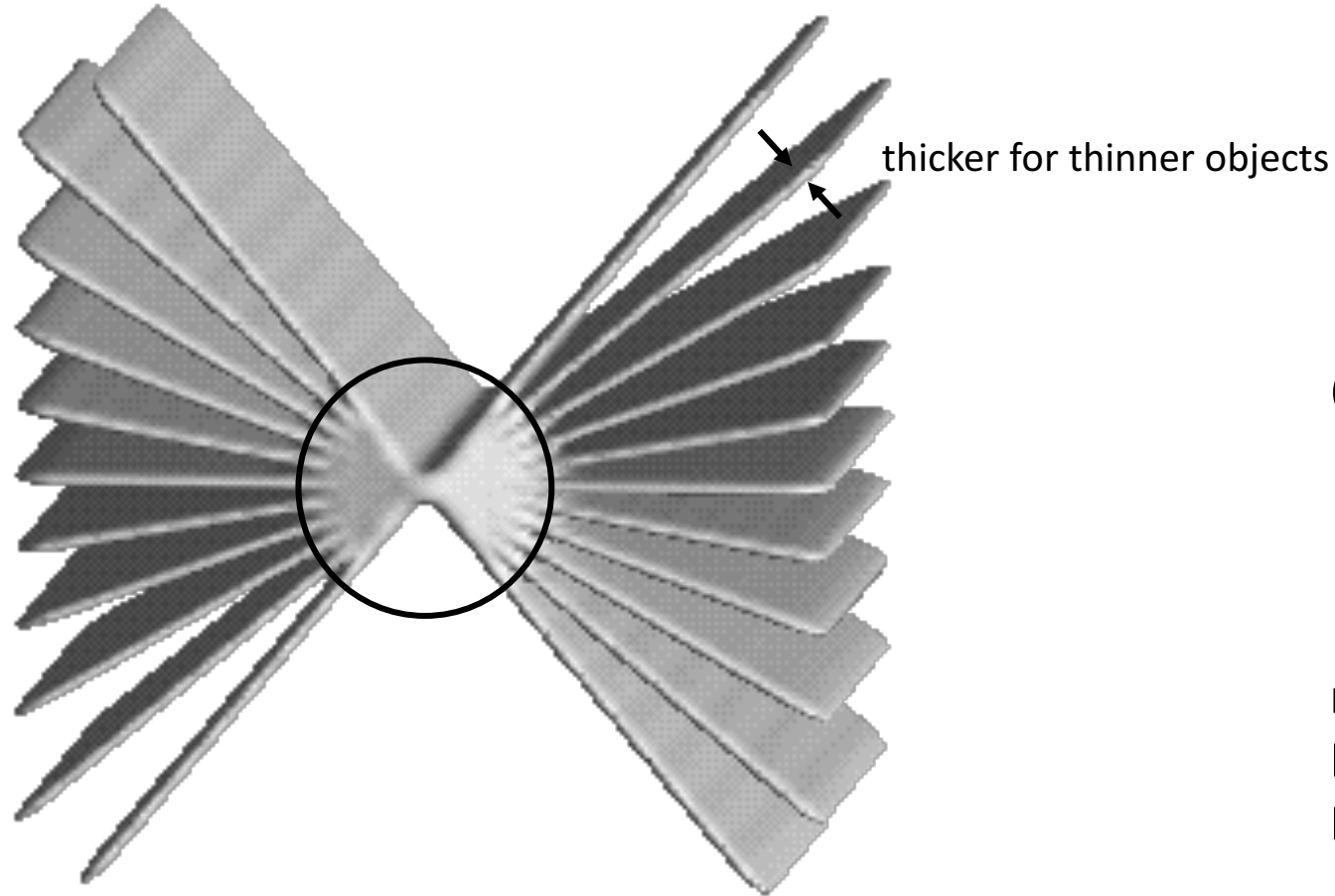
2D projections -> 3D reconstruction



# The missing wedge leads to smearing in z



# Missing information due to discrete sampling

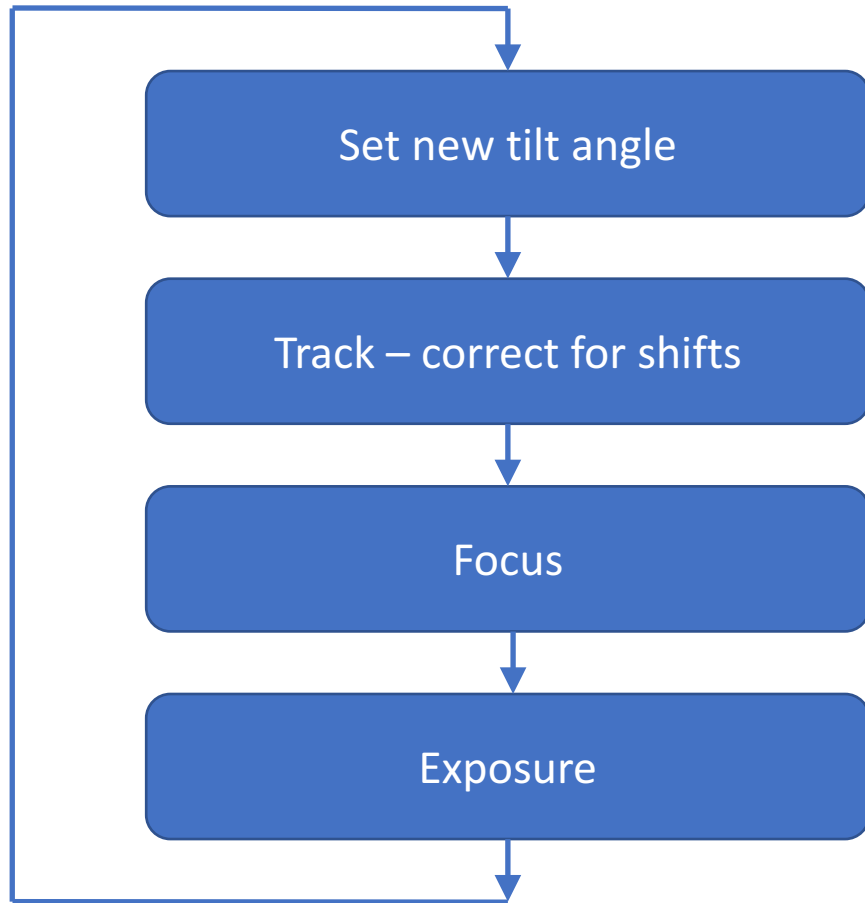


Crowther Criterion

$$r = \pi D / N$$

r: Resolution limit  
N: Number of projections  
D: Object thickness

# Data collection



What questions should we ask before data collection?

# Data collection

What total dose? Resolution vs signal-to-noise

What tilt range? Completeness of information vs dose and speed

What angular increment? Resolution vs dose and speed

What order to collect the images? Speed, reliability, optimal dose, sample distortion...

What magnification? Resolution and DQE vs field of view

What defocus? High-frequency information vs low frequency information



# Alignment and Reconstruction

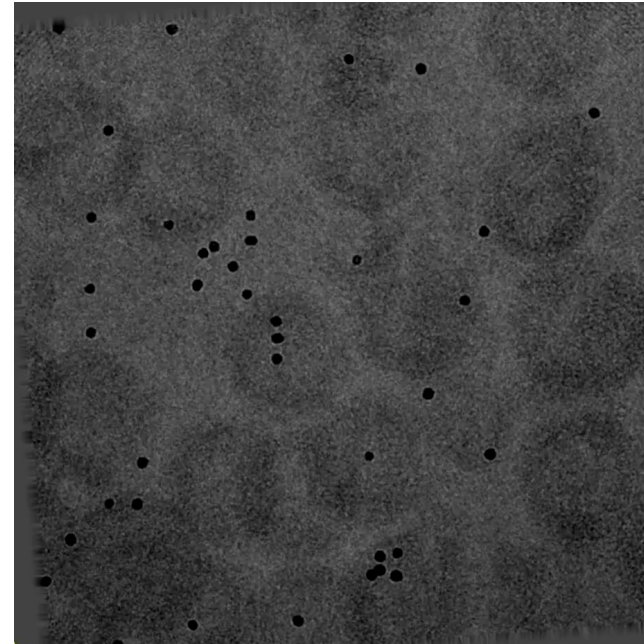
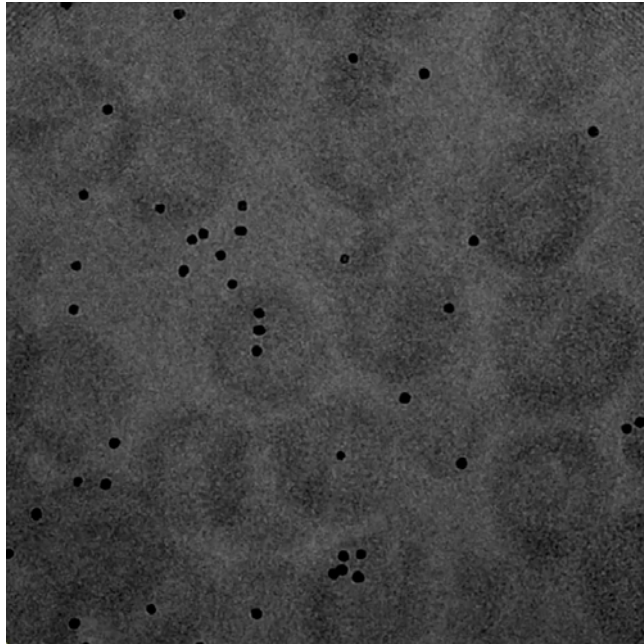
Once we have collected the data how do we reconstruct a tomogram?

# Alignment and Reconstruction

We need to know how the projections relate to each other: the angles and shifts between the projections.

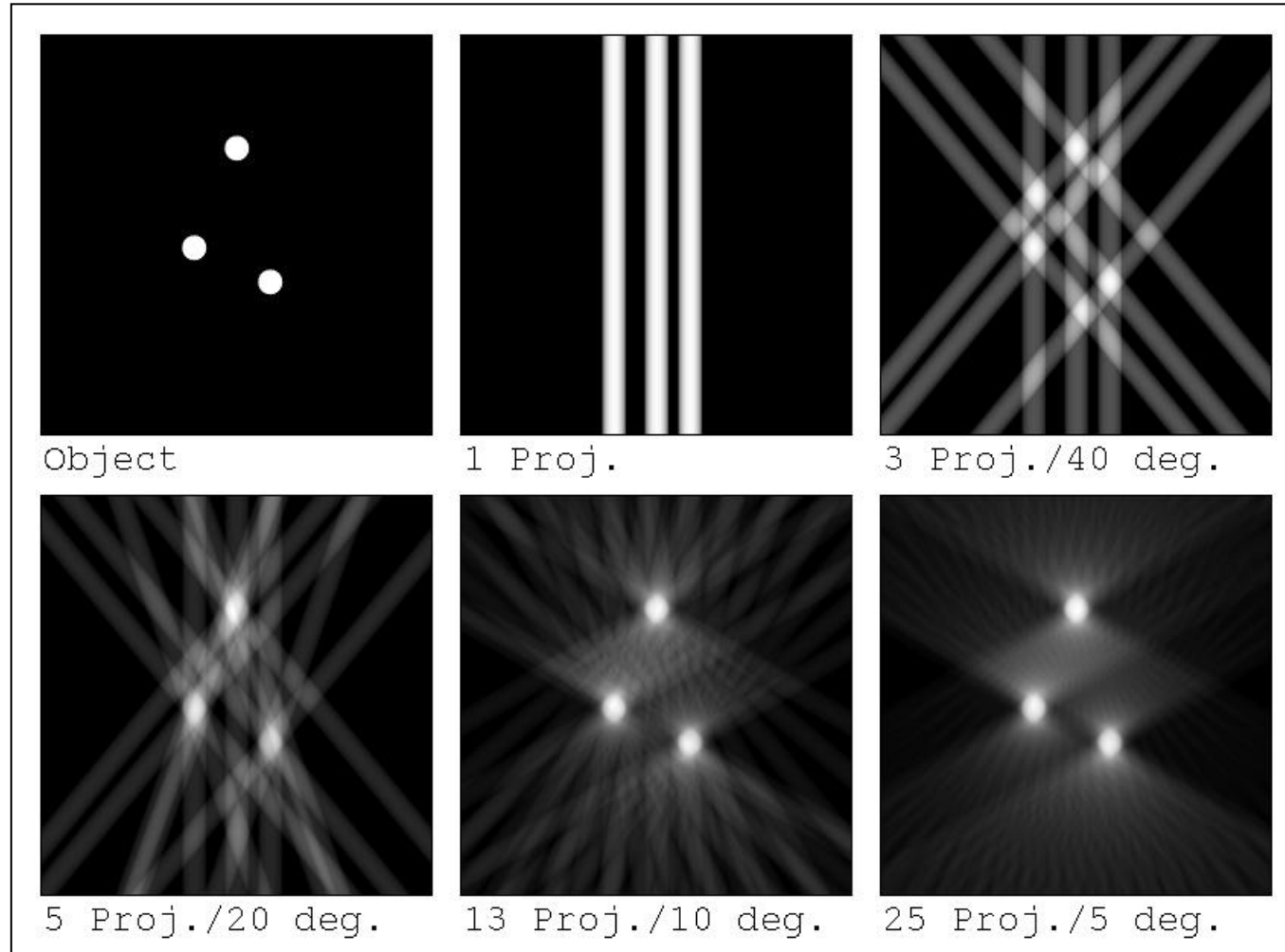
We have defined the angles in the microscope by tilting around a defined axis by defined increments.

Alignment is necessary to deal with the shifts in the image.  
(at larger fields of view, other distortions may become important)

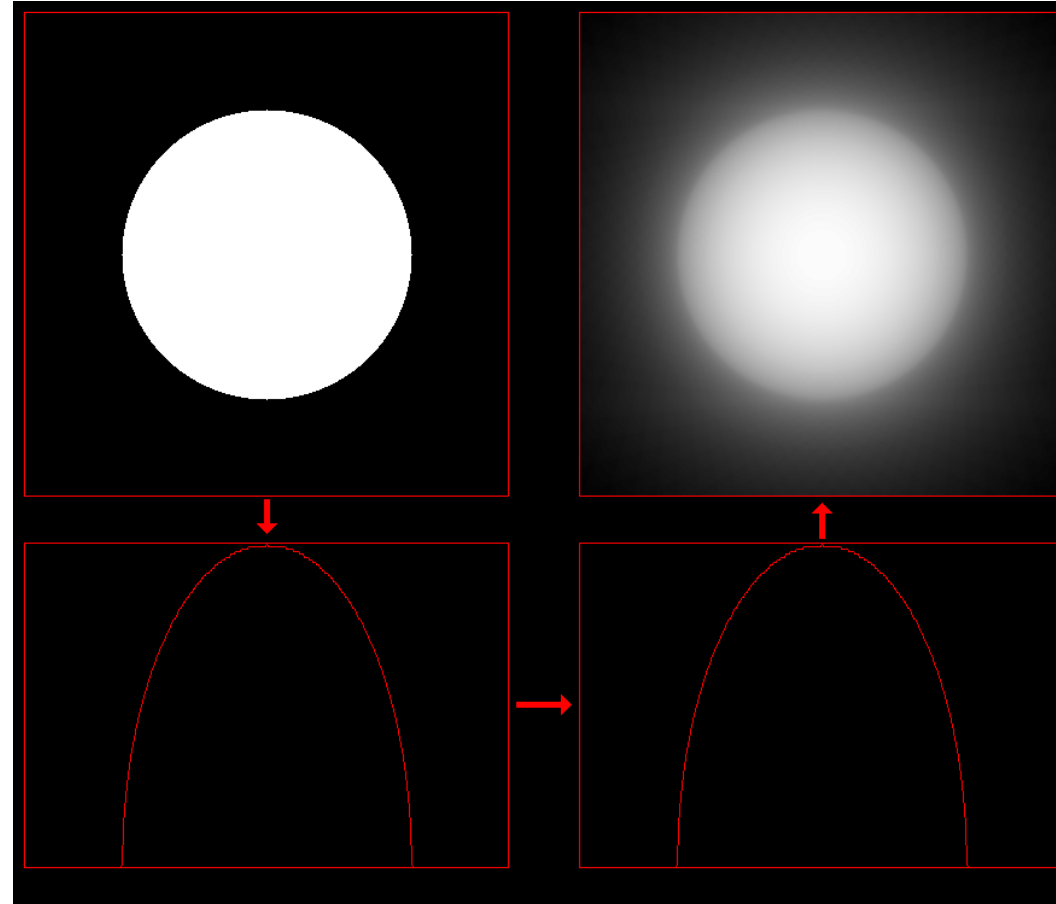


# Alignment and Reconstruction

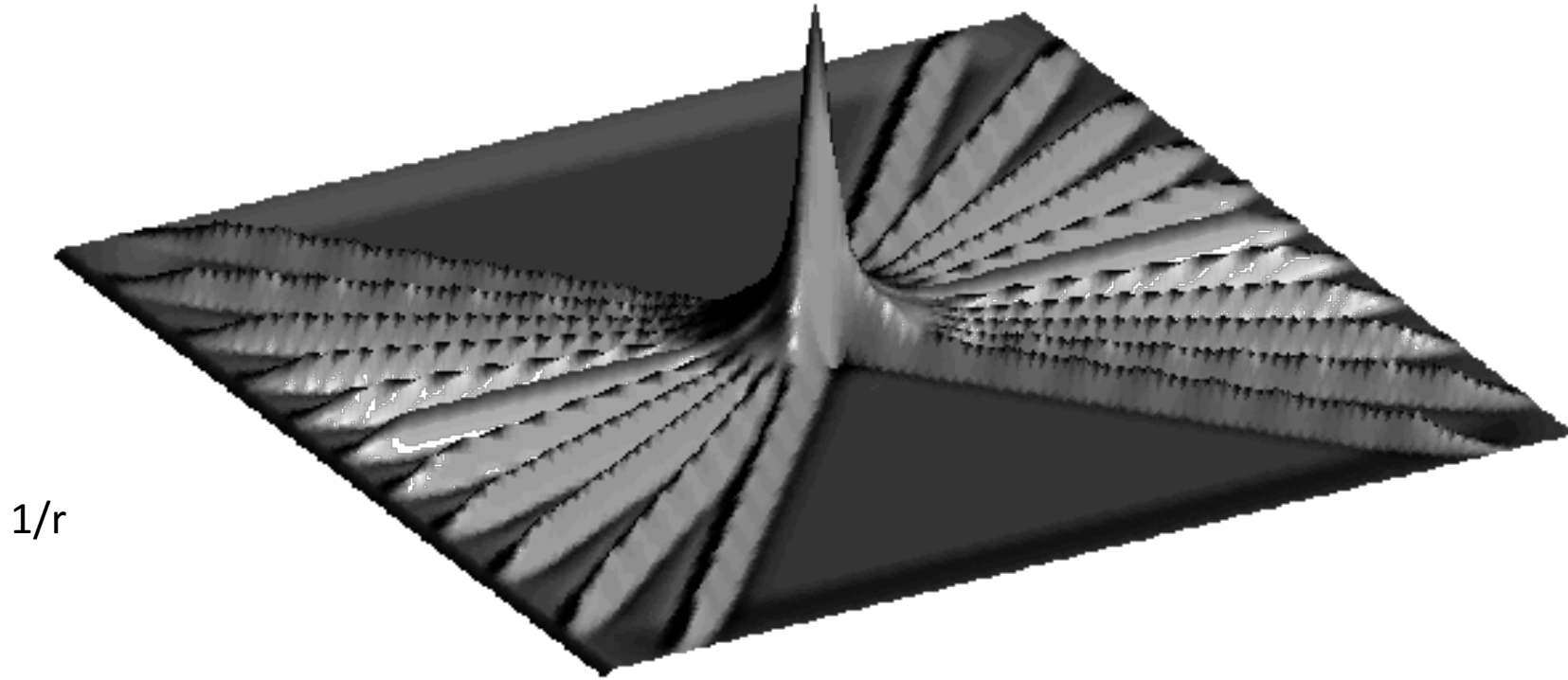
- Reconstruction by weighted back projection



# Back-projection

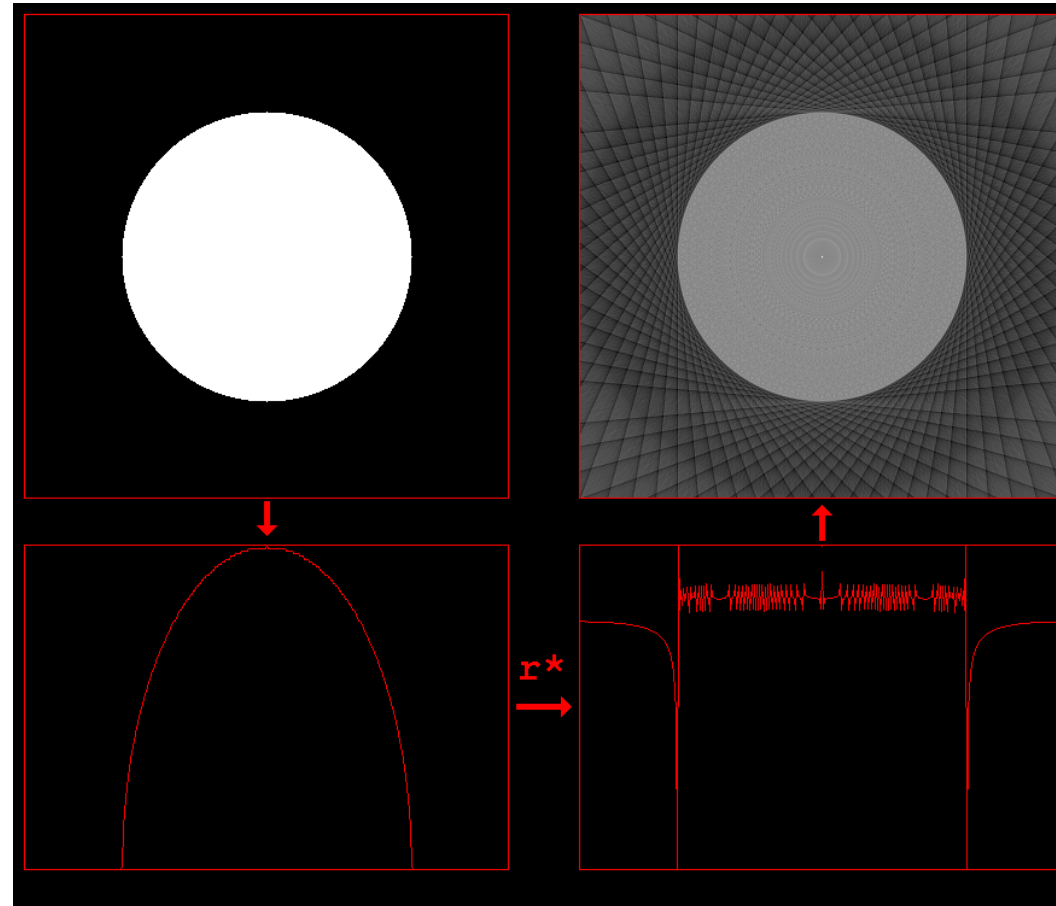


# Back-projection



$1/r$

# Back-projection



# Software

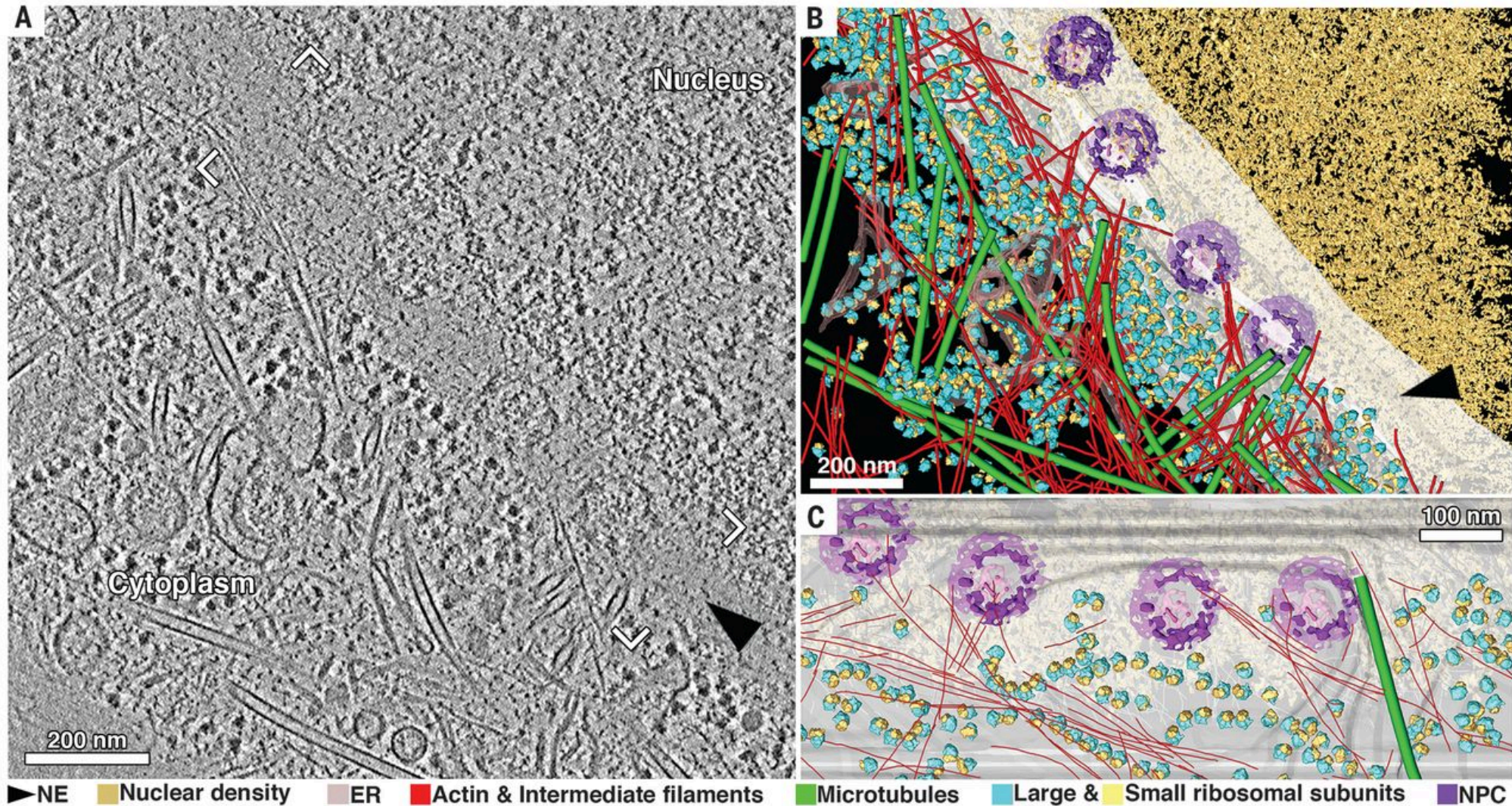
Data Collection: **SerialEM**, FEI Tomo, Leginon...

Tomogram reconstruction: **IMOD**, TOM, protomo, PyTOM...

# Subtomogram averaging

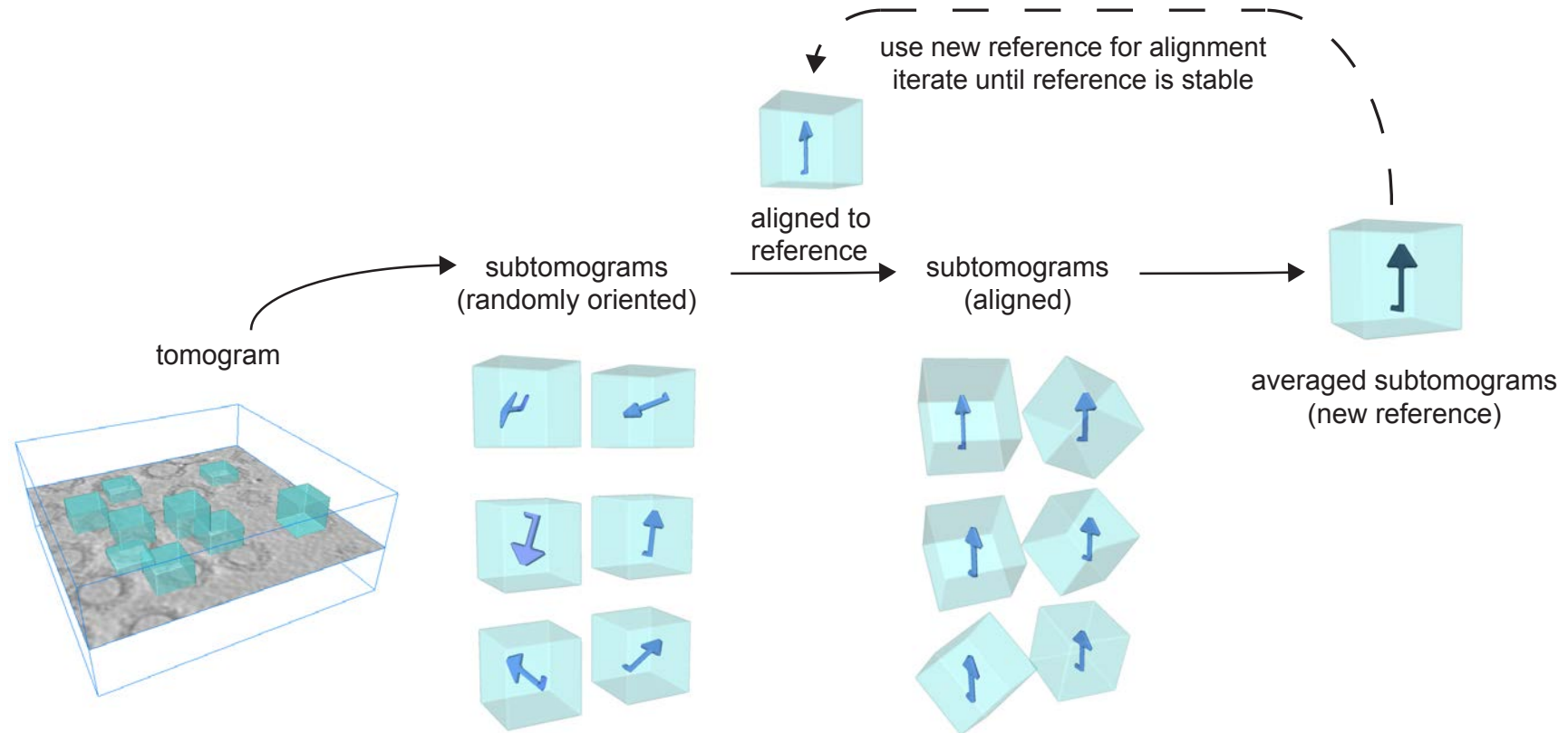


# Subtomogram averaging

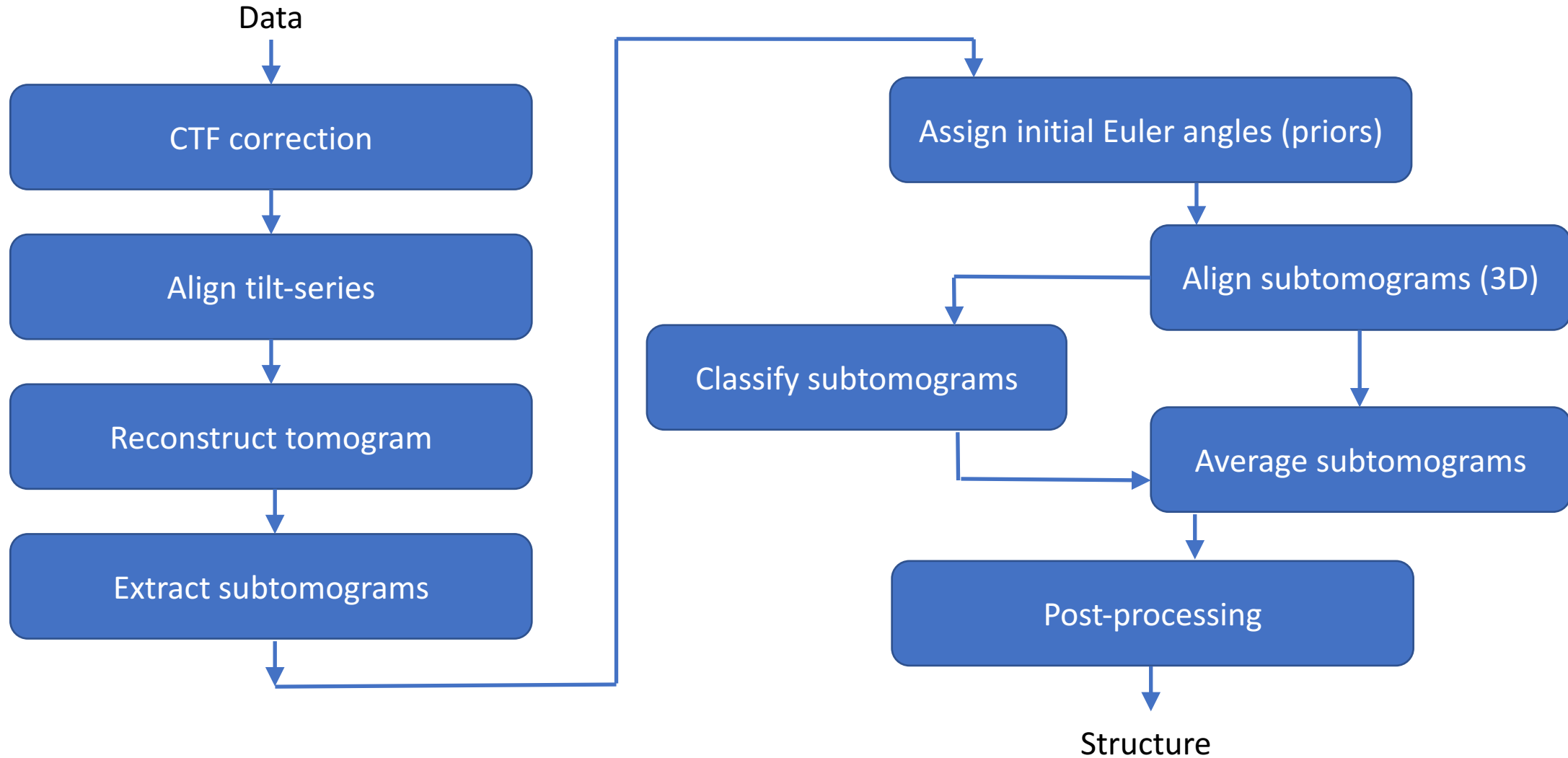




# Subtomogram averaging



# Subtomogram averaging process



# Software for subtomogram averaging

Dynamo (Castano-Diez, Basel)

PEET (Heumann and Mastronarde, Boulder)

PyTOM (Foerster, Utrecht)

RELION (Bharat and Scheres, LMB)

Maximum cross-correlation or Maximum likelihood

# Subtomogram averaging

Why not just do single particle reconstruction? (if you can, then do it!)

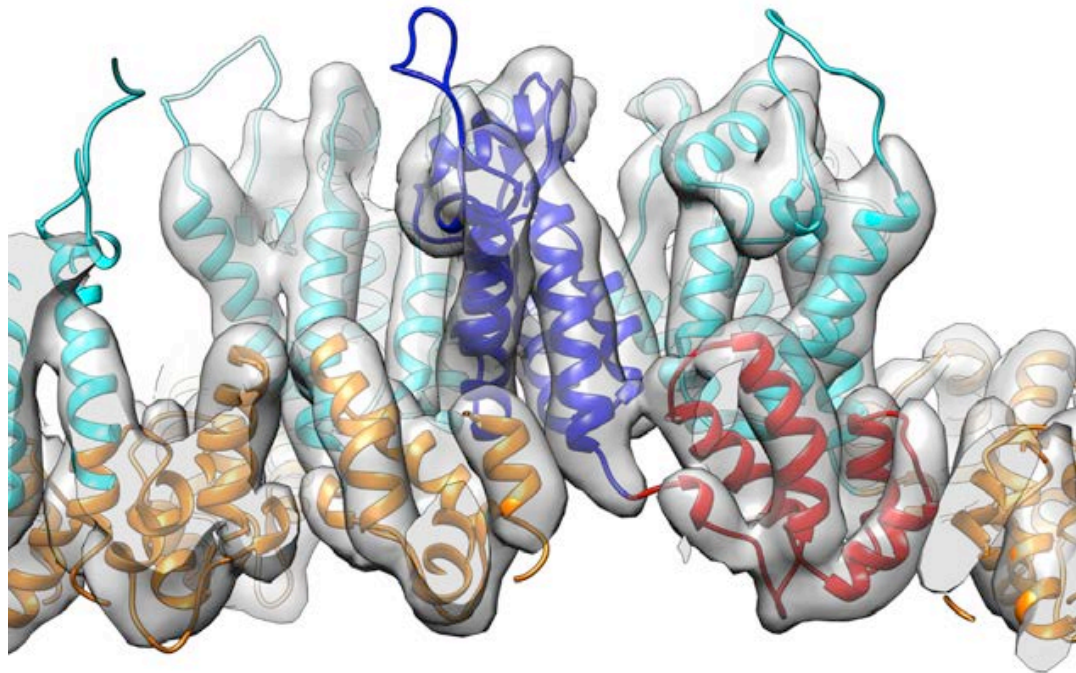
Subtomogram averaging allows structures to be determined when other objects in the path of the electron beam would otherwise prevent alignment.

(and has potential for application in single-particle type projects)

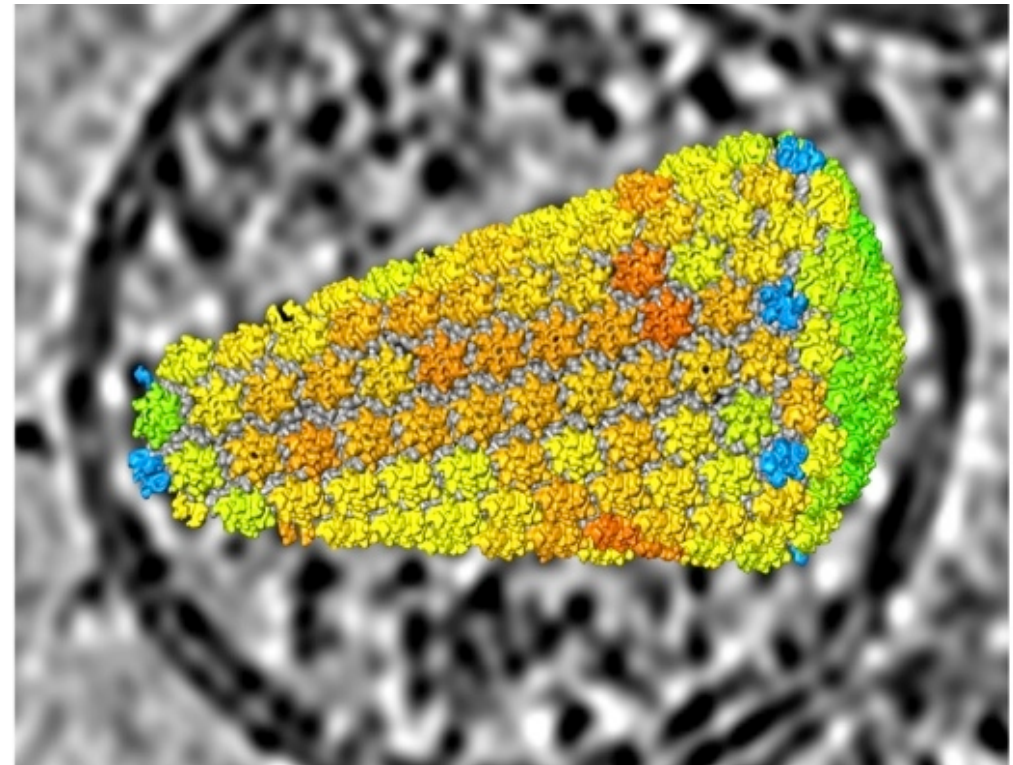
# Subtomogram averaging

Can generate two kinds of information

Structure



Position and context



# Subtomogram averaging - challenges

Sample flexibility and heterogeneity

Higher apparent sample thickness (especially at tilt)

Two alignment and reconstruction steps

Sample movement/change during data collection

Higher total electron dose

Smaller datasets (due to more time-consuming data collection and processing)

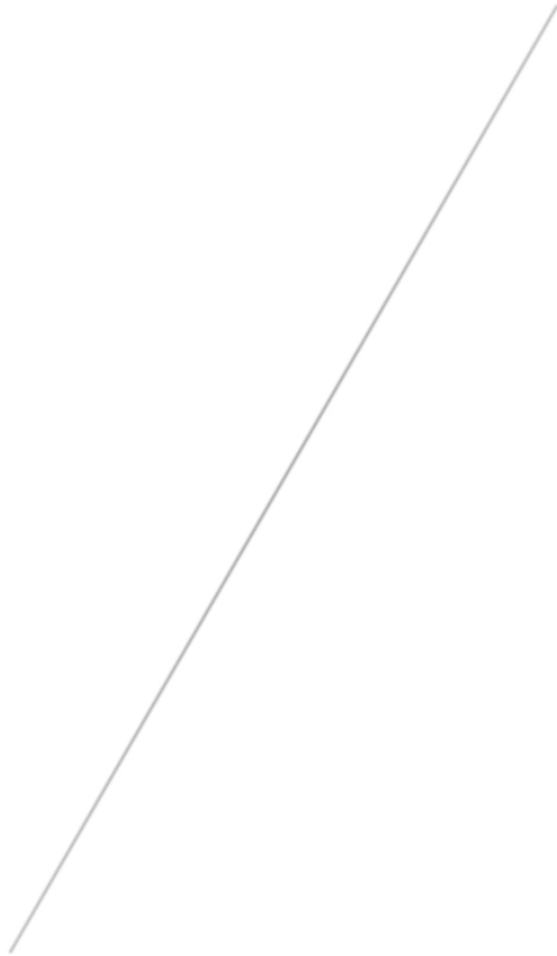
Difficult in determination and correction of CTF

# Subtomogram averaging - challenges

In which order should we collect the images?



# Continuous tilt scheme



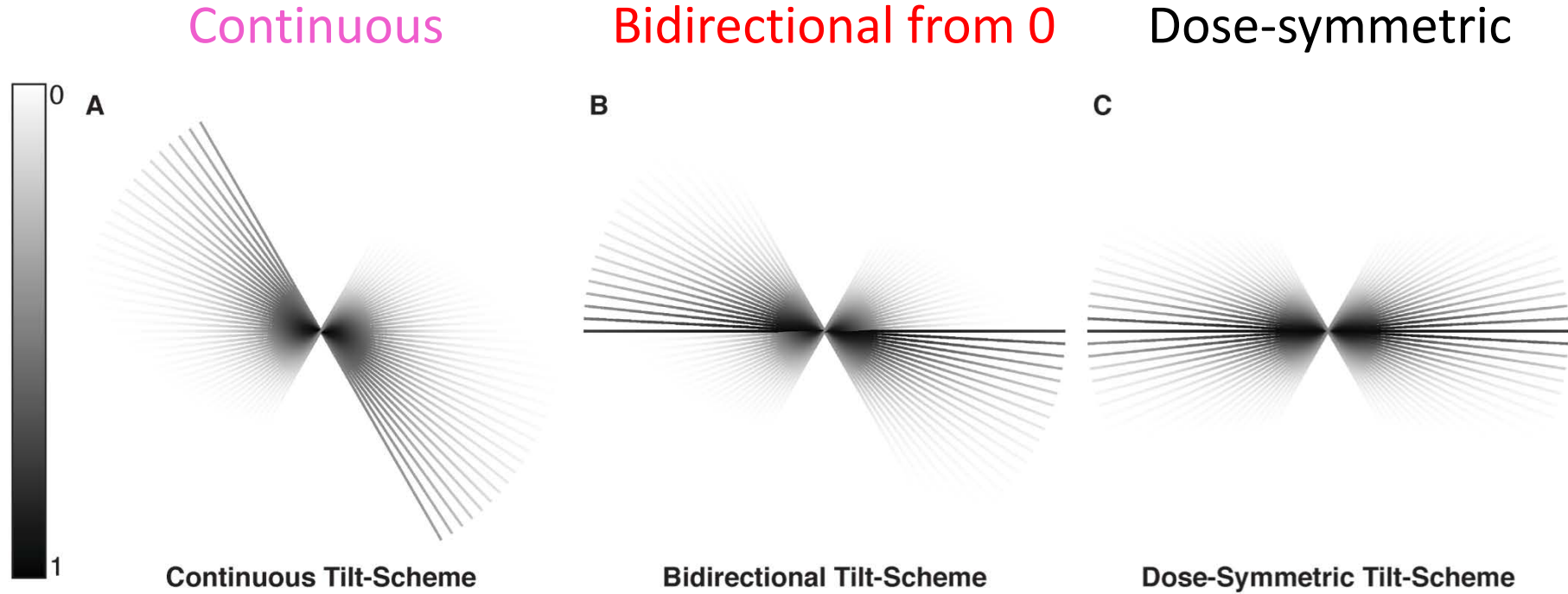
# Bidirectional tilt scheme



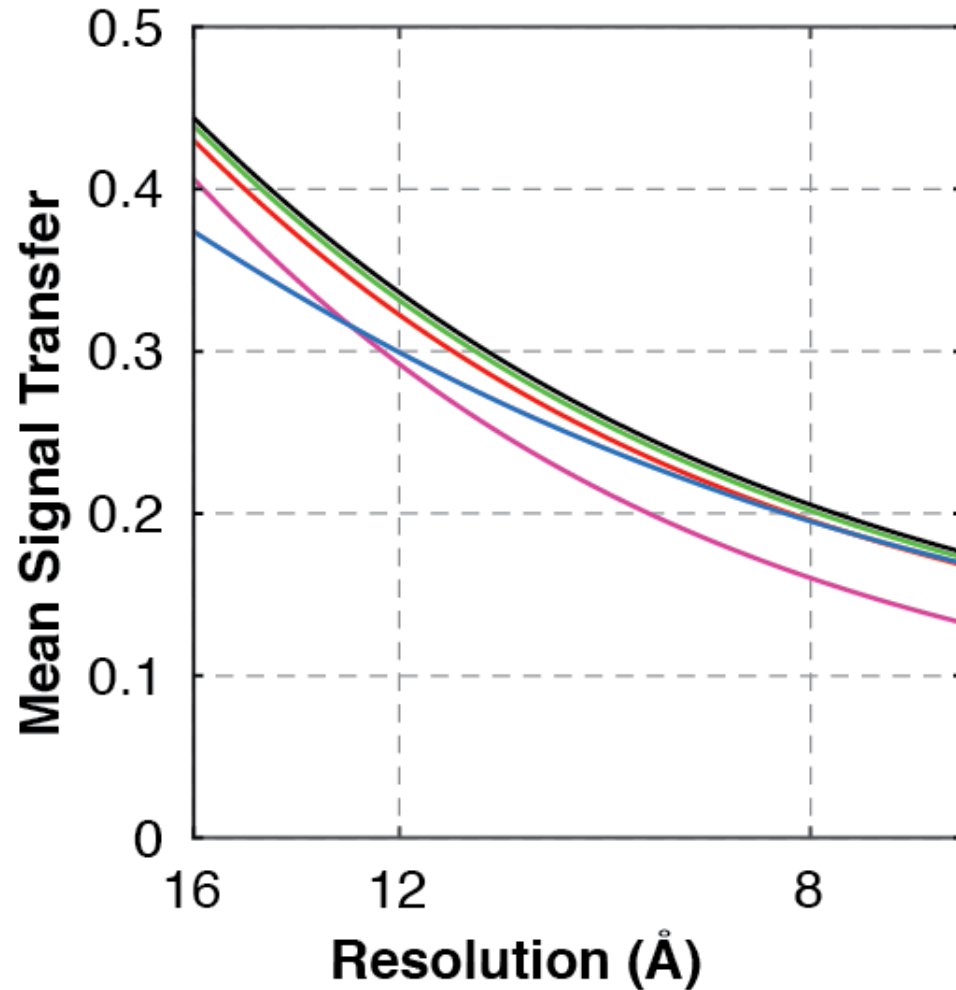
# Dose-symmetric tilt scheme



# Tilt schemes



# Tilt schemes – signal transfer



Continuous

Bidirectional from 0

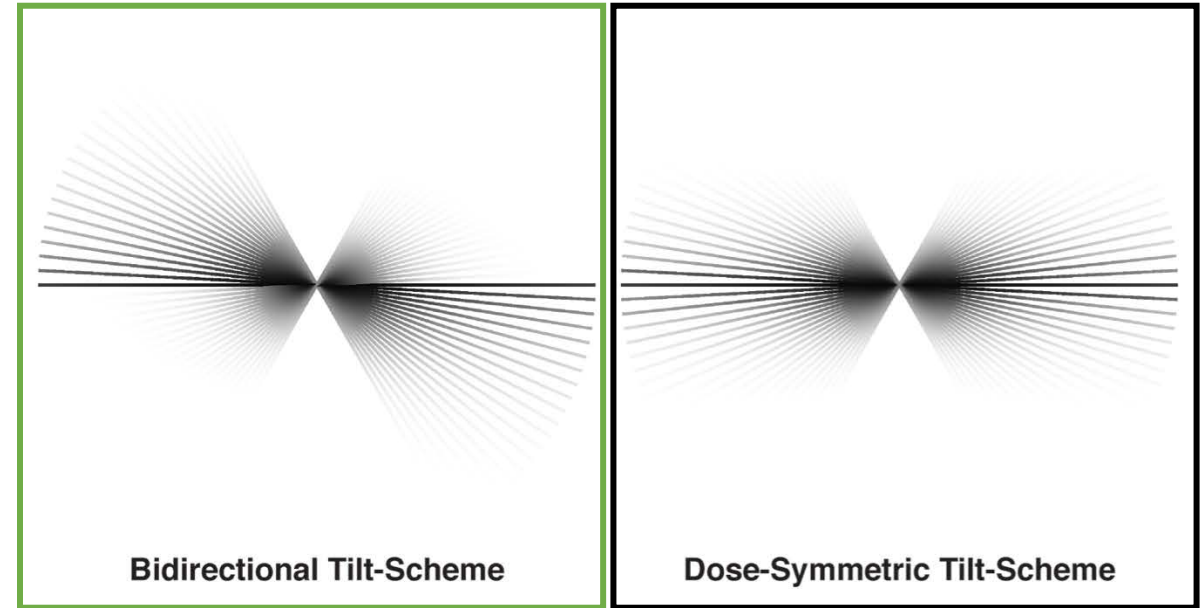
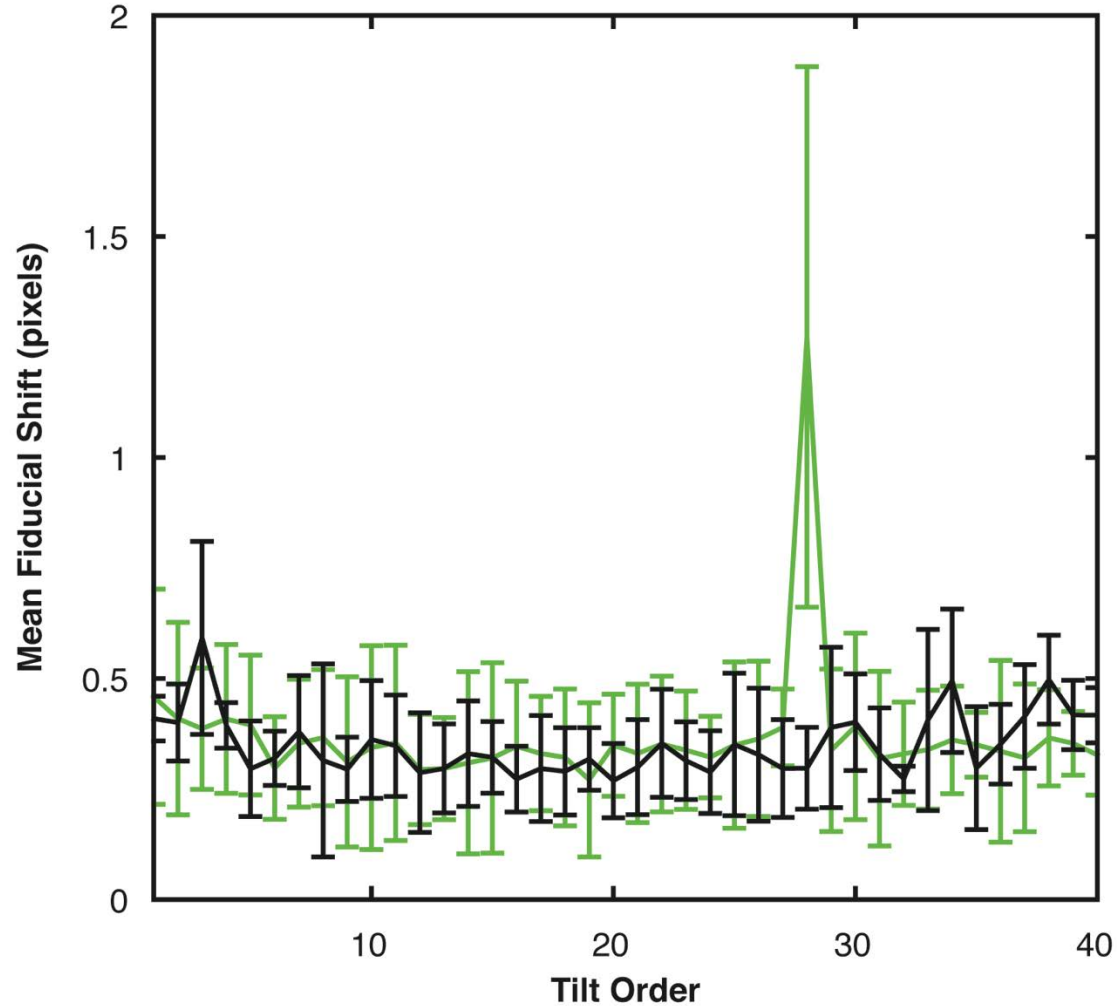
Bidirectional from -21

Bidirectional from -21  
deleting second branch

Dose-symmetric

Note – the difference is greater if you also consider increased sample movement at tilt

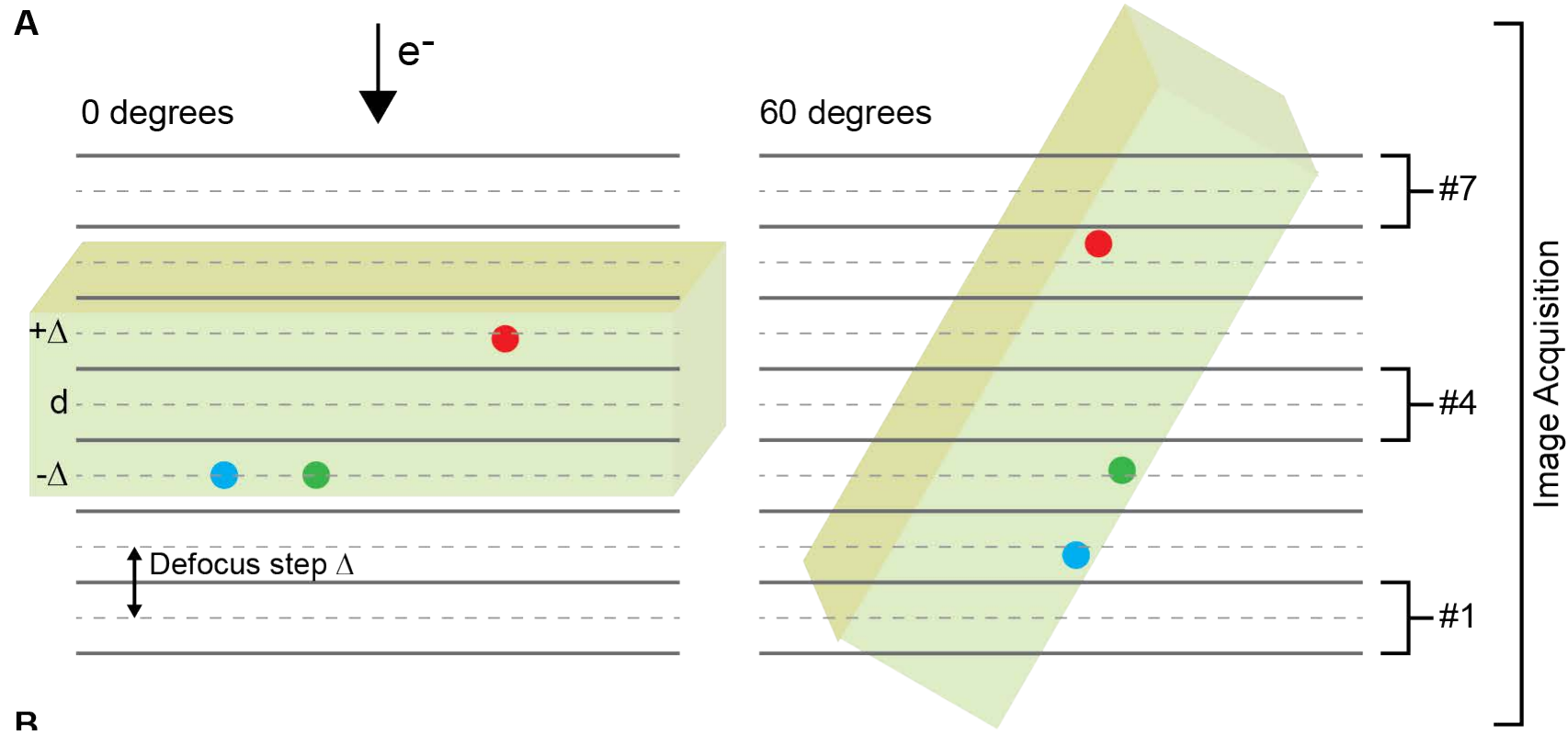
# Dose-dependent sample changes



# Subtomogram averaging - challenges

CTF Correction

# Defocus gradients in the sample

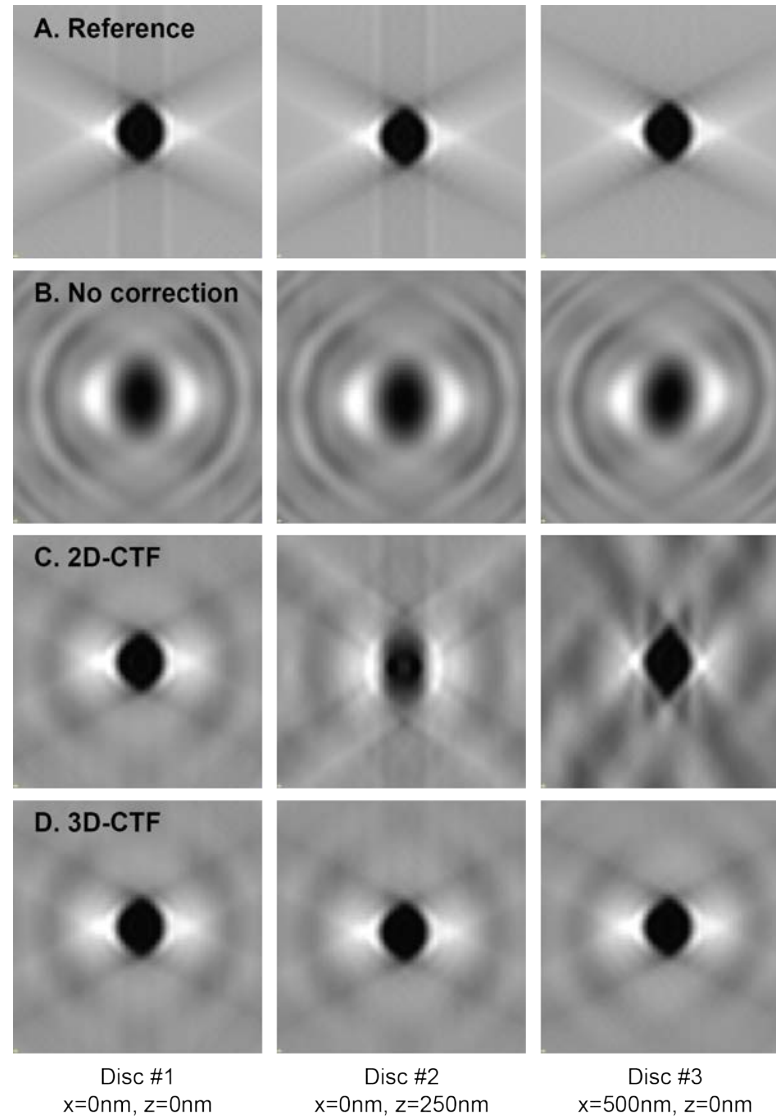


Turonova. et al

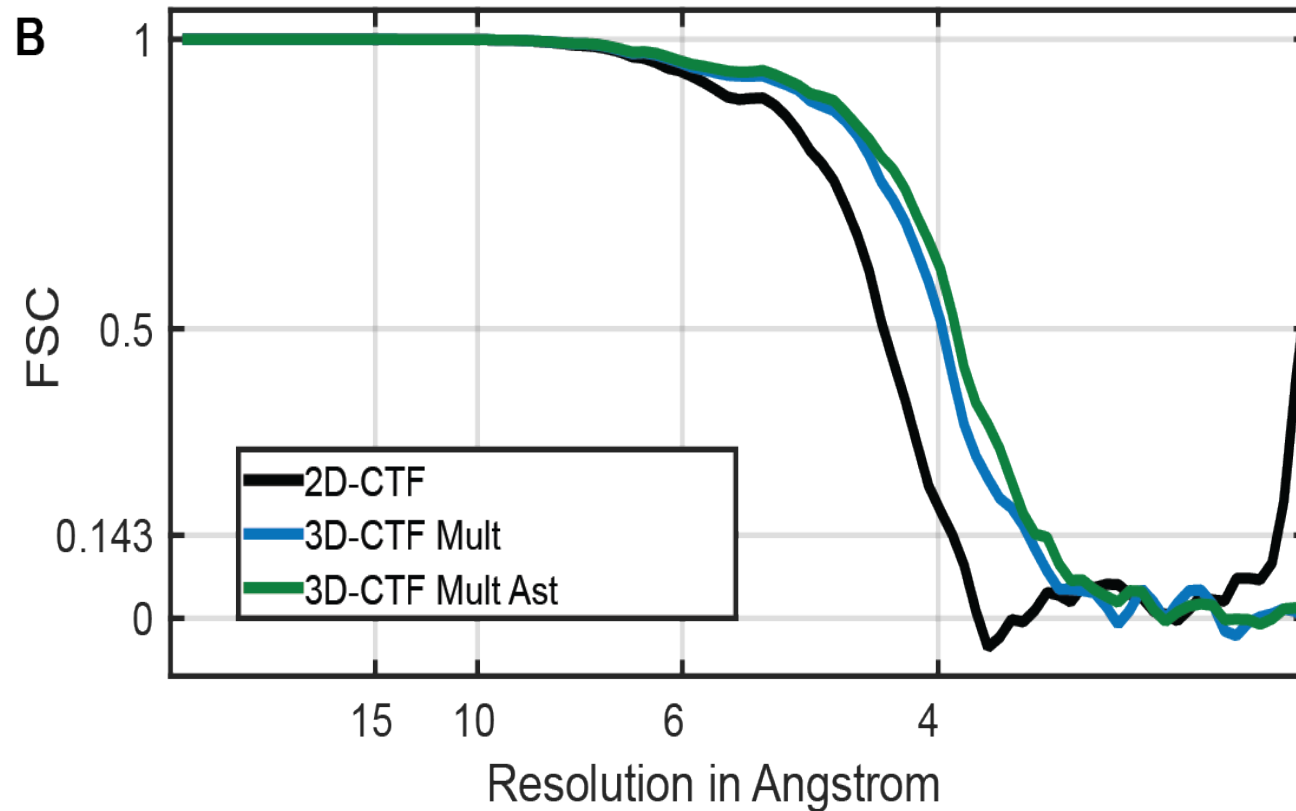
2D CTF correction considers only the gradient due to tilt, 3D CTF also considers the gradient through the thick sample



# Defocus gradients in the sample



# Defocus gradients in the sample



# Subtomogram averaging - challenges

Sample flexibility and heterogeneity

Higher apparent sample thickness (especially at tilt)

Separate alignment and reconstruction steps

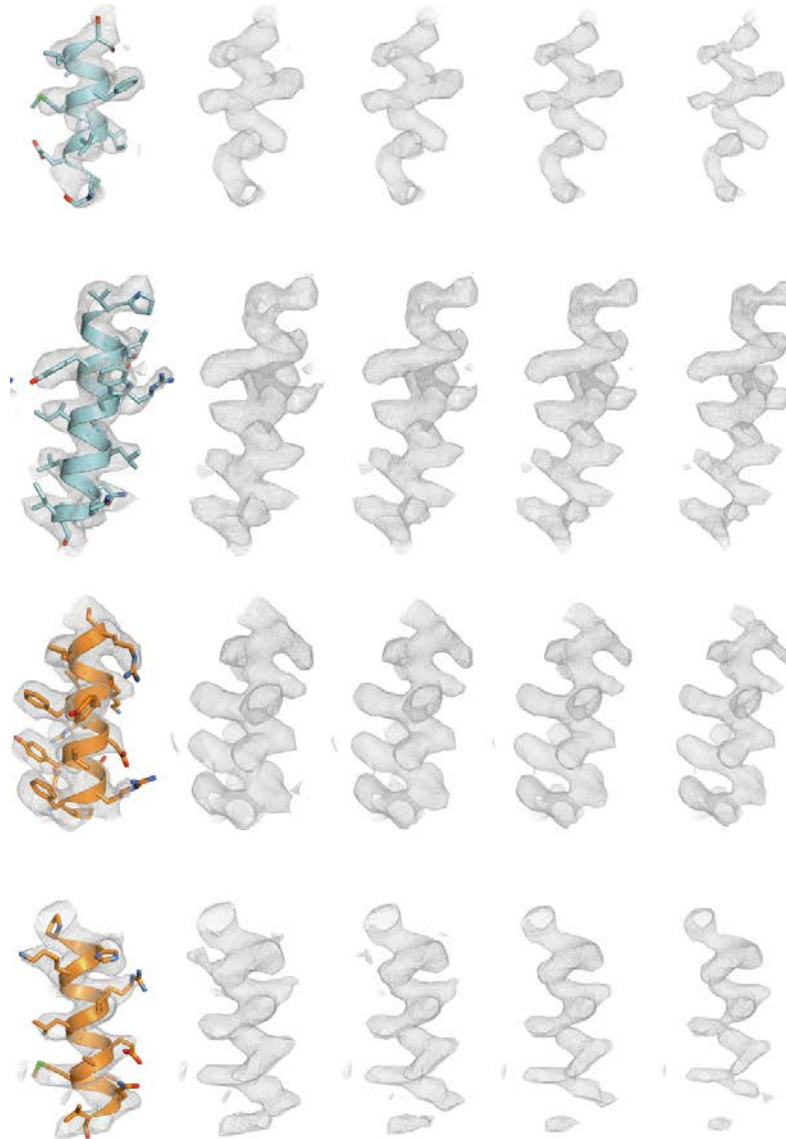
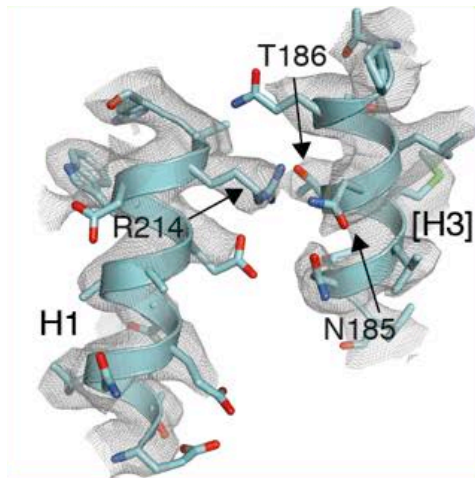
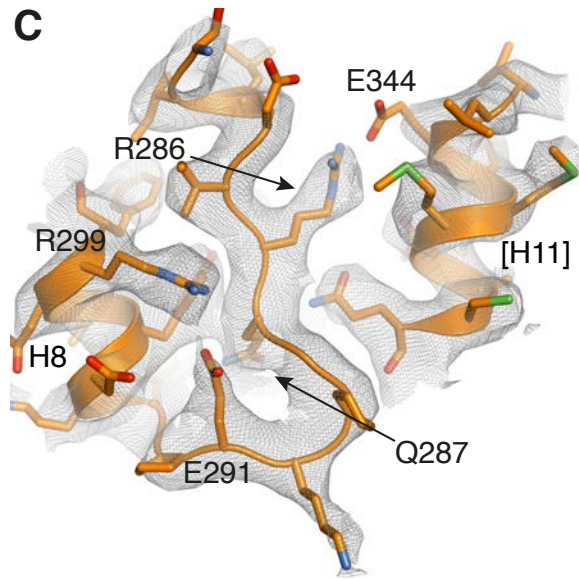
Sample movement/change during data collection

Higher total electron dose

Smaller datasets (due to more time-consuming data collection and processing)

Difficult in determination and correction of CTF

# It is possible to obtain $<4\text{\AA}$ resolution





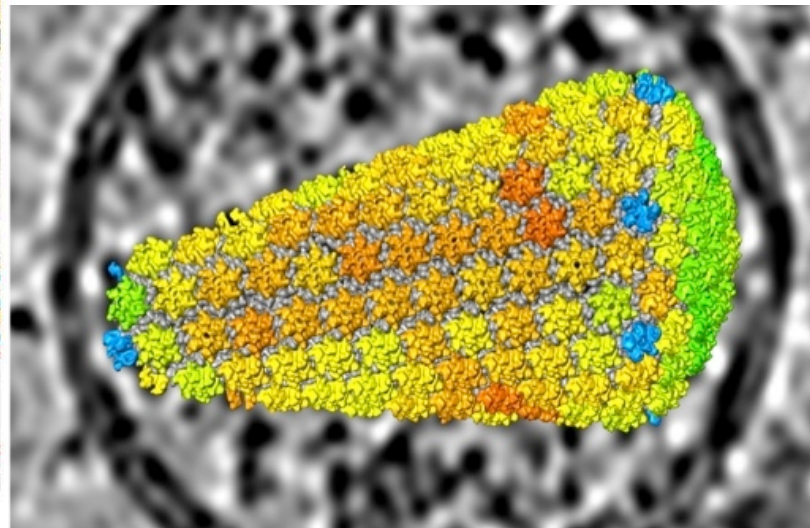
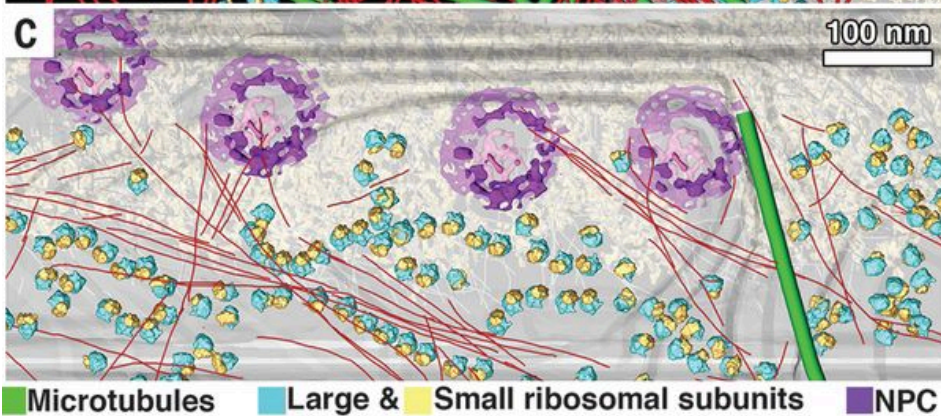
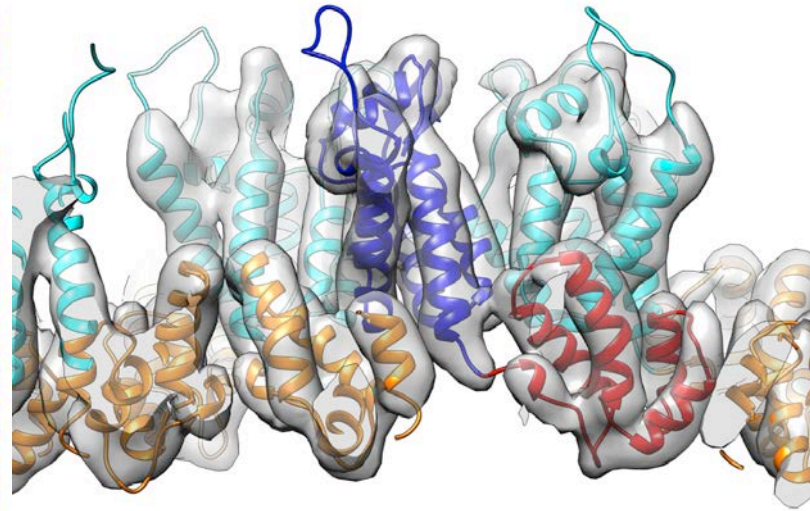
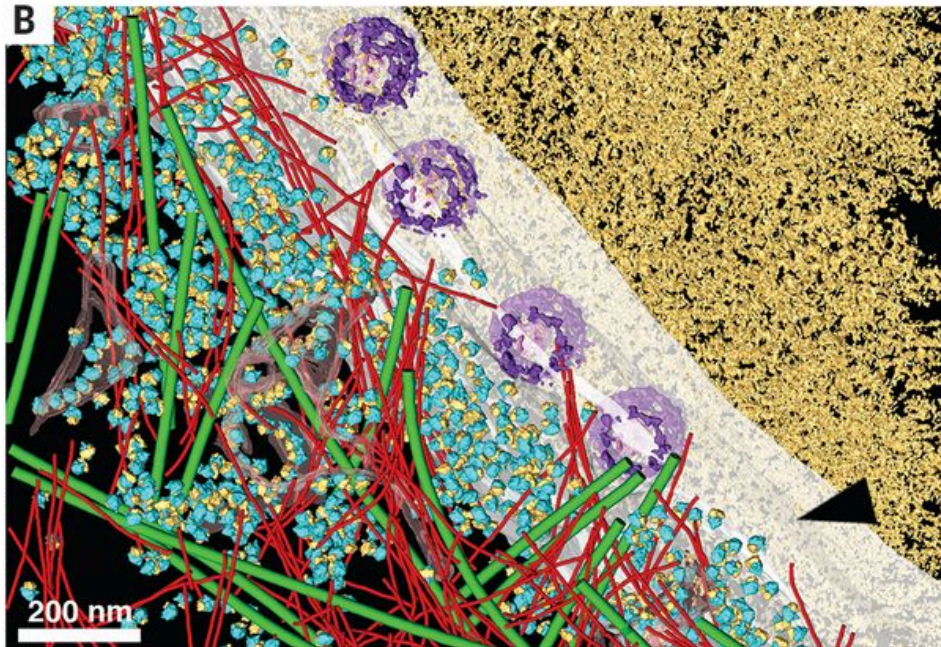
# Subtomogram averaging

Structure “in situ”

Cellular context

Structure for  
“Discovery”

Potential SP  
applications



Wan, W. and Briggs, J.A.G., “Cryo-electron tomography and subtomogram averaging”  
(2016) *Methods in Enzymology*, 579, 329-367

and articles by many authors cited therein