### Model-Building with Coot

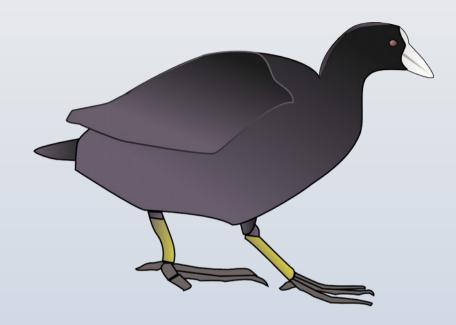
#### Adding N-linked Glycosylation & Demo

Paul Emsley MRC Laboratory of Molecular Biology May 2017

### About this Presentation

- Modelling N-linked Carbohydrate
- Demo: Carbohydrate Fitting
- Demo: Protein-fitting
  - [Coot & Key-bindings]
- When is the available?
  - Available from the *Coot* Web Site now
    - Get the latest 0.8.9-pre pre-release
  - much is in 0.8.8
  - And CCP4
- pdf

### Coot Collaborators

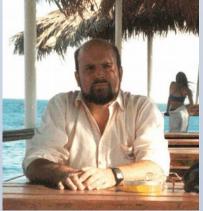




Bernhard Lohkamp



Kevin Cowtan









Richardsons & co-workers, Duke

Eugene Krissinel Stuart McNicholas Martin Noble

Alexei Vagin

### A Brief History of Coot

- Released in 2004, Coot was designed primarily for model-building protein models into maps from x-ray data
  - Torsions: Rotamers, Ramachandran plots
  - Several optimisers, including Real Space Refinement
- Used typically after automated model-building or refinement
- Since:
  - Nucleic Acids, Ligands & Cryo-EM



- It's never been pretty...
  - Not the best tool for presentation graphics and animations

### Coot Key-bindings

- Many hundreds of functions available in Coot's API
  - available via scheme or python
- Coot's gui doesn't help much to learn key-bindings
  - they are "off" by default
  - so that you can program your own
- If you are more than a casual/occasional users of *Coot...* are probably worth learning

#### Using *Coot* on a Mac

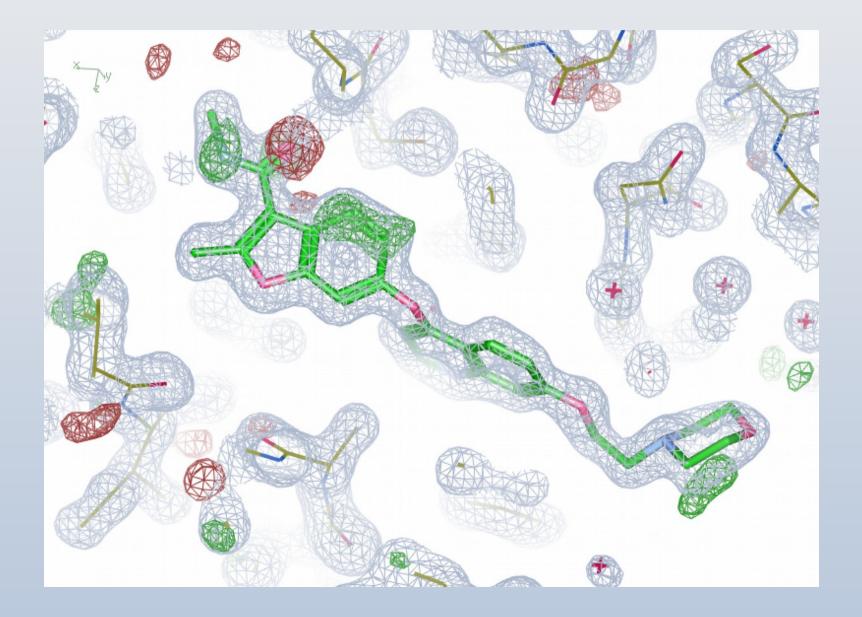
- Coot is an X11-based application
- Xquartz  $\rightarrow$  Preferences
  - Input
    - Emulate 3-button Mouse
  - Windows
    - Focus Follows Mouse

### Making Density Slides with Coot

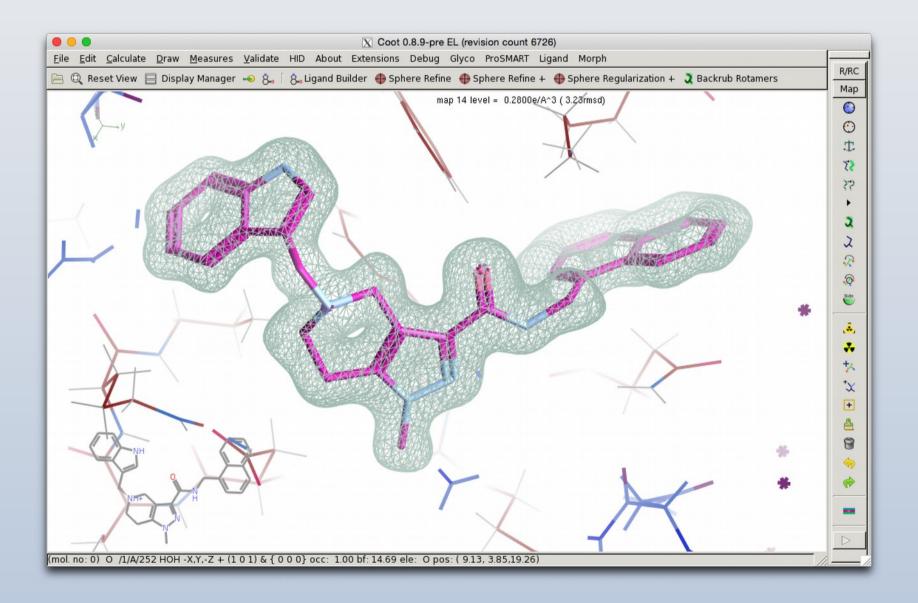
- White background
- "High" Oversampling (2.3x)
- Pale gray (or very pastel) density colour
- Enable Cut-glass mode 5-10%
- Licorice Bonds (Shift-4)
  - Highlight Interesting Site
- Anti-aliased Coot

- \$ setenv \_\_GL\_FSAA\_MODE 5

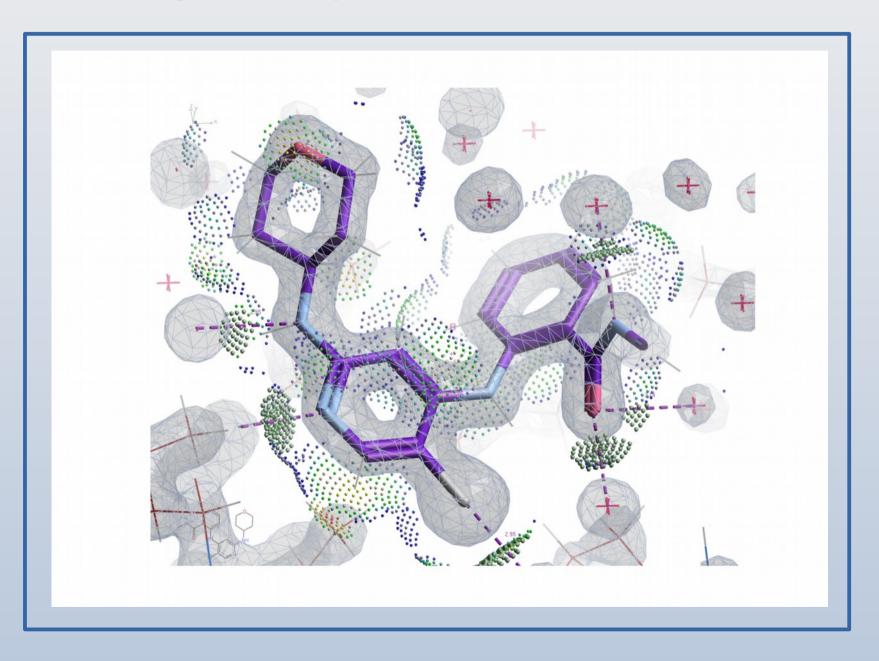
### Example Density Slide



#### Ligands Representation in Coot



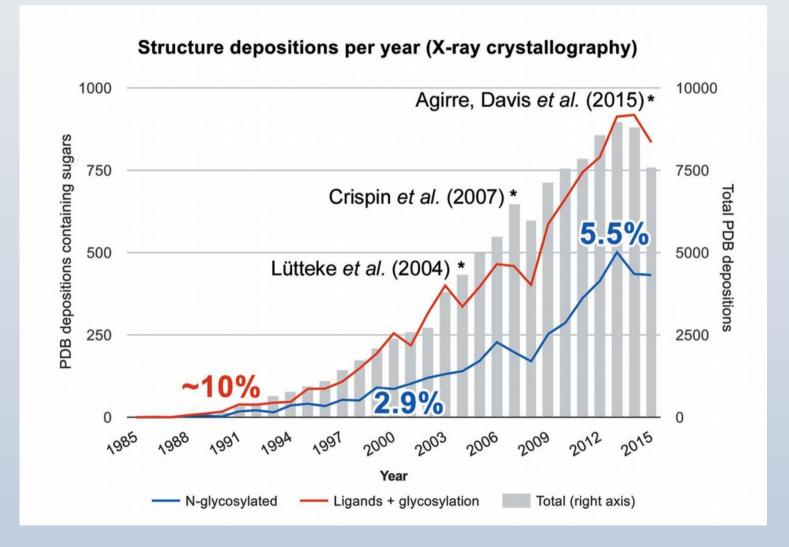
#### Ligand Representation in Coot



### Glycoproteins

- Glycosylation is the most frequent and most complex protein modification
- Unlike protein, the structure is glycans is only indirectly encoded in the genome
  - (glycosyltransferases)
- Even in the same cell, a particular type of protein can be differently glycosylated
  - microheterogeneity
- Although N-linked glycans are relatively conserved
- Glycans stabilize the protein and shield it from attack by proteases
- Play a role in cell-cell signalling:
  - fertilization, differentiation, host-pathogen, immune response

#### Glycoproteins

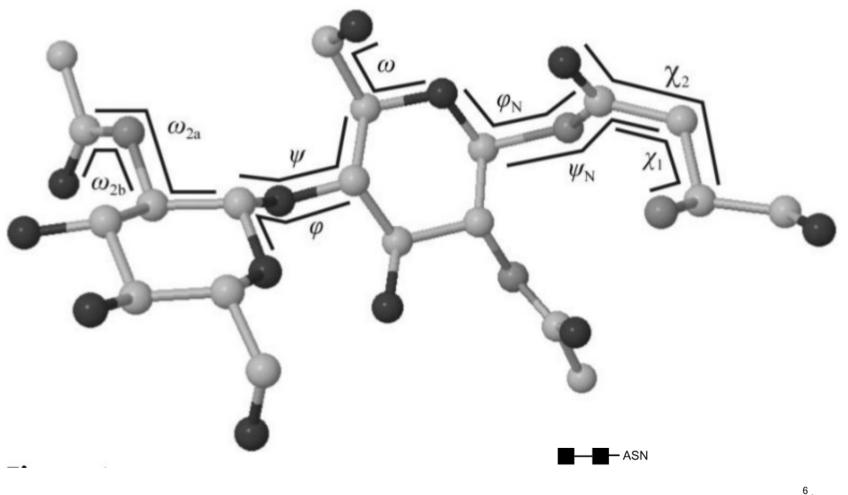


Agirre (2017)

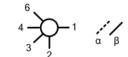
### Problematic Glycoproteins

- Crispin, Stuart & Jones (2007)
  - NSB Correspondence
  - "one third of entries contain significant errors in carbohydrate stereochemistry..."
  - "carbohydrate-specific building and validation tools capable of guiding and construction of biologically relevant stereochemically accurate models should be integrated into popular crystallographic software. Rigorous treatment of the structural biology of glycosylation can only enhance the analysis of glycoproteins and our understanding of their function"
  - PDB curators concur
  - More recently Joosten & Lűtteke (2017), Agirre et al. (2017)

### **Carbohydrate Links**

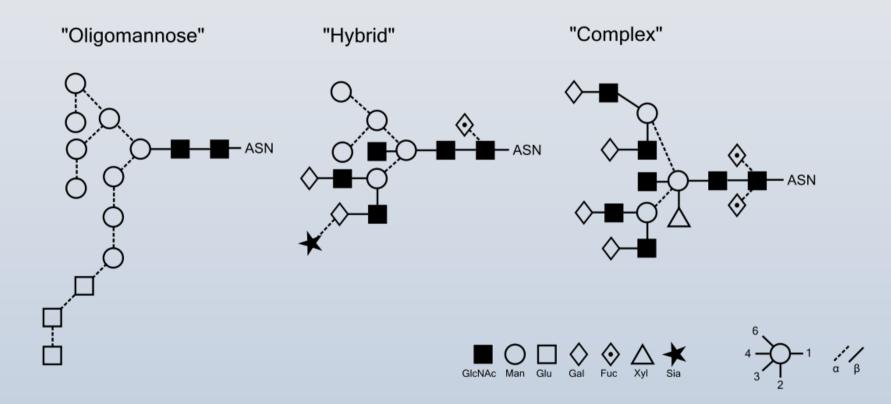






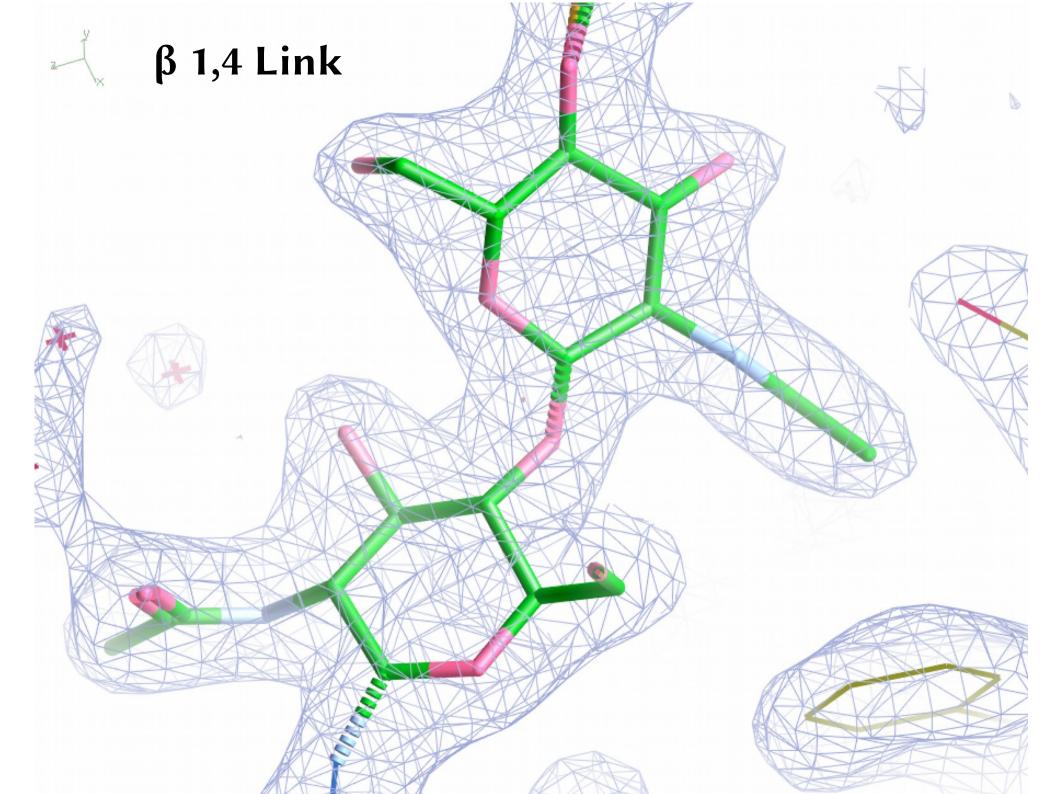
Thomas Lütteke (2007)

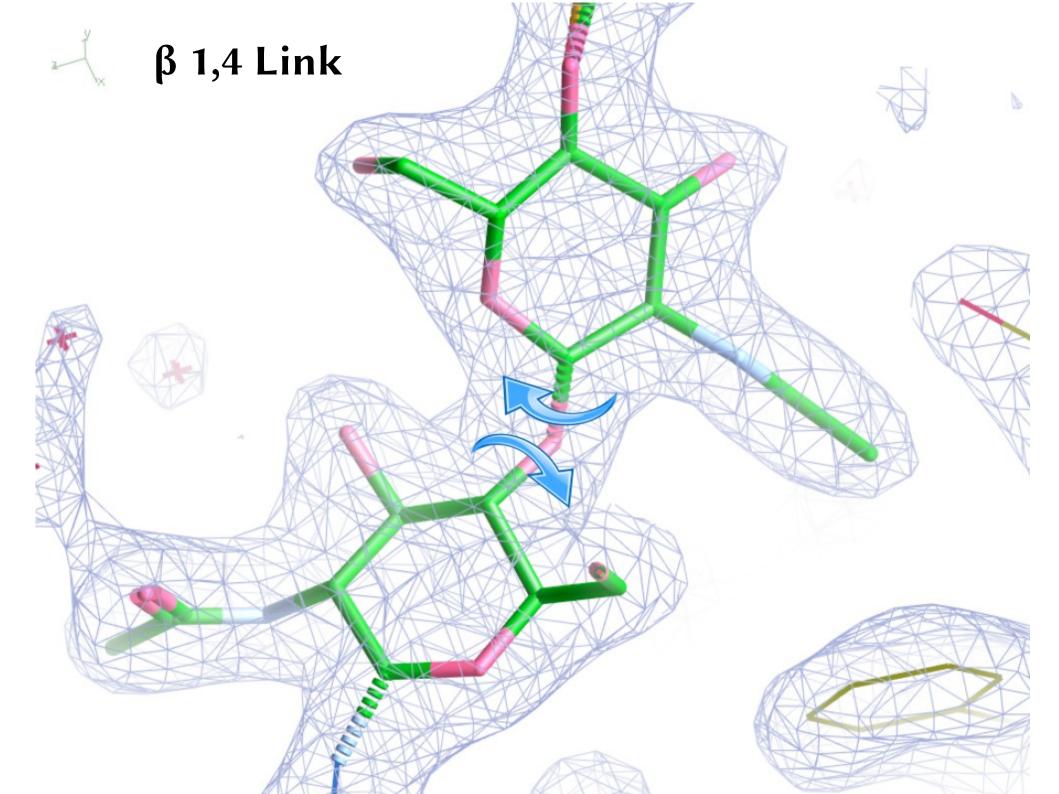
#### N-linked carbohydrates

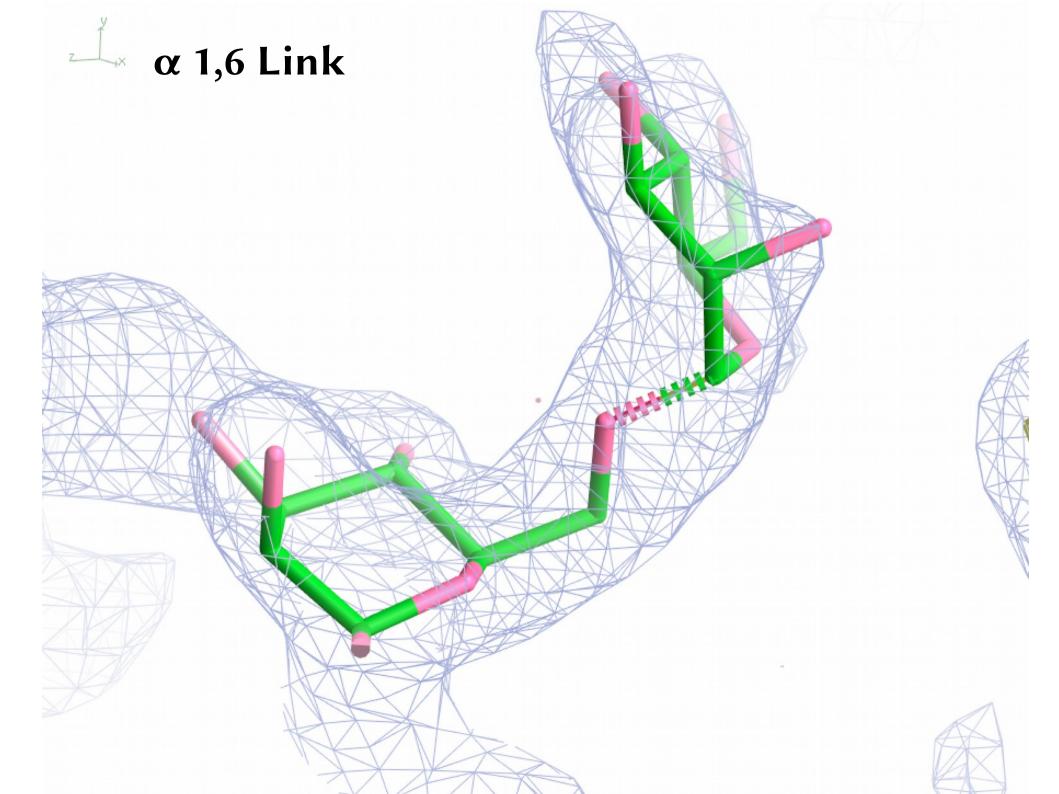


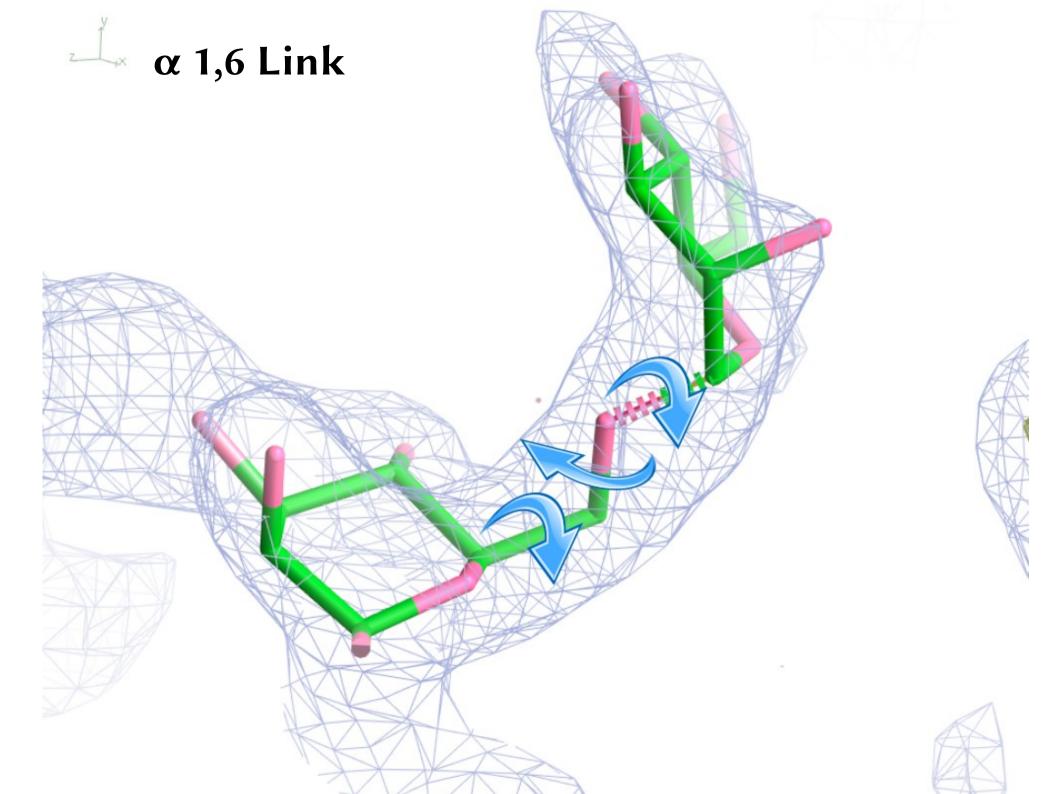
### Linking Oligosaccharides/Carbohydrates: LO/Carb

- One can fully define carbohydrate structure by the primary structure and a set of torsion angles
- Build complex carbohydrate structure
  - from a dictionary of standard links
  - and monomers
  - torsion-angle refinement
    - by simulated annealing

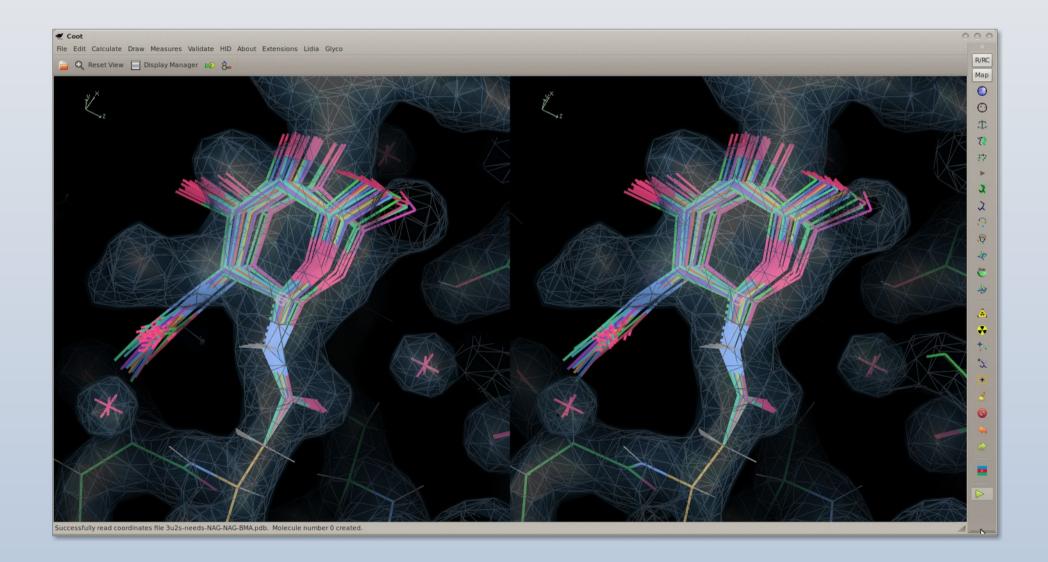








# Refinement Progress (NAG-ASN example)

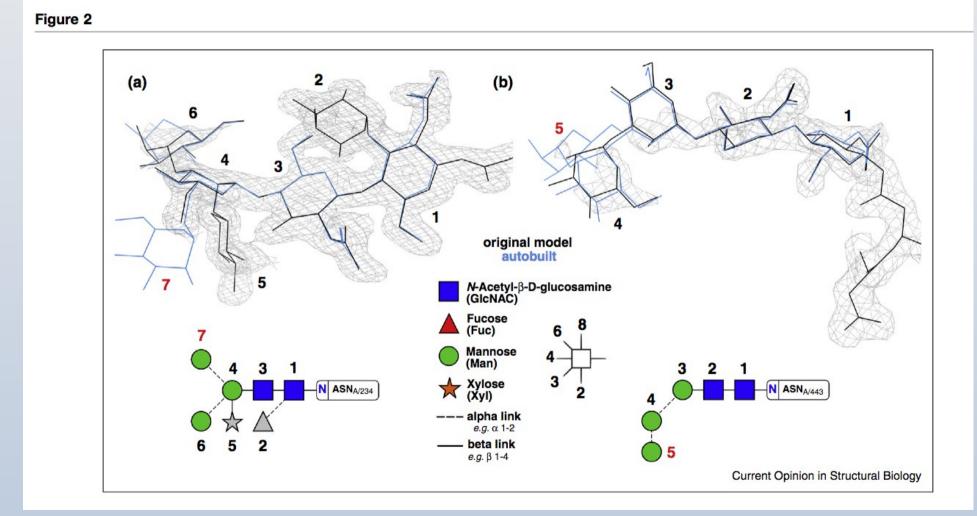


### N-linked Glycan Modelling in Coot

- Two Modes:
- Residue by Residue
  User Control
- Whole Tree Addition
  - Automated
  - Embedded decision-making

Glyco	Restraints					
Add	a ASN-NAG NAG					
Add a BETA1-4 NAG						
Add a BETA1-4 BMA						
Add an ALPHA1-2 MAN						
Add	an ALPHA1-3 MAN					
Add	an ALPHA2-3 MAN					
	an ALPHA2-3 GAL					
Add	an ALPHA1-6 MAN					
	a BETA1-2 NAG					
Add	a BETA1-4 GAL					
	an ALPHA1-3 FUC					
	an ALPHA1-6 FUC					
	an BETA1-6 FUL					
	an XYP-BMA XYP					
	k add NAG, NAG, BMA					
	Oligomannose					
Add Paucimannose						
Add Complex Tree						
Delete All Carbohydrate						
Torsion Fit this residue						
Torsion Fit & Refine this residue						
Add synthetic pyranose plane restraints						
Use Unimodal ring torsion restraints						
Glyco Tree Residue ID						
N-trials to 500						
N-trials to 1500						
N-tri	ials to 5000					

### Problematic Glycosylation



#### Agirre et al. (2017) The Rocky Road to Automation

14/40

### Linking Fucose: Fuc-α1,3

- Add a menu item to wrap the command
  - add\_linked\_residue("FUC", "ALPHA1-3")

Added into a new N-linked tree:

- paucimannose

## Xyl- $\beta$ 1,2

- Xyl β1,2 Man
  - using XYP (beta D xylosepyranose)
  - was not in the Refmac Monomer Library list of links
  - It has been added and will be available to CCP4 shortly

### **Refinement Stabilizers**

- Using Coot's Real Space Refinement
  - (in default mode)
  - allowed saccharides to result in twisted or boat ring conformations
    - even without user intervention
- Unimodal Torsion Restraints
- Pseudo-plane restraints
- Inter-residue Geman-McClure external distance restraints

### REFMAC Monomer Library chem\_comp\_bond

loop\_

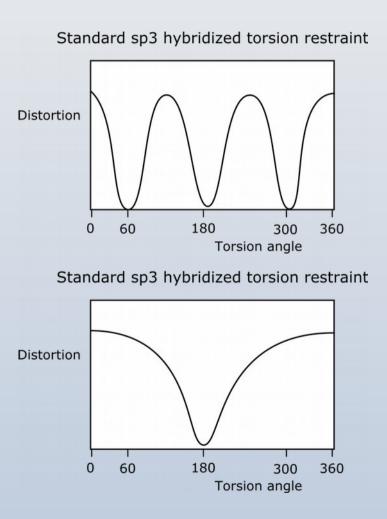
_chem_comp_bond.comp_id					
_chem_comp_bond.atom_id_1					
_chem_co	omp_bond	d.atom_id	_2		
_chem_co	omp_bond	d.type			
_chem_co	omp_bond	d.value_d	ist		
_chem_co	omp_bond	d.value_d	ist_esd		
ALA	Ν	Н	single	0.860	0.020
ALA	Ν	CA	single	1.458	0.019
ALA	CA	HA	single	0.980	0.020
ALA	CA	СВ	single	1.521	0.020
ALA	СВ	HB1	single	0.960	0.020
ALA	СВ	HB2	single	0.960	0.020

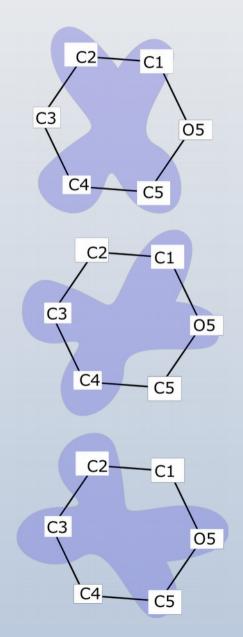
### REFMAC Monomer Library chem\_comp\_tor

loop_						
_chem_comp_tor.comp_id						
_chem_c	omp_tor.id					
_chem_c	omp_tor.at	om_id_1				
_chem_c	omp_tor.at	om_id_2	)			
_chem_c	omp_tor.at	om_id_3	}			
_chem_c	omp_tor.at	om_id_4				
_chem_c	omp_tor.va	lue_ang	le			
_chem_comp_tor.value_angle_esd						
_chem_c	omp_tor.pe	riod				
ADP	var_1	02A	PA	C		
	var 2	D٨	037	C		

ADP	var_1	02A	PA	03A	PB	60.005	20.000	1
ADP	var_2	PA	03A	PB	01B	59.979	20.000	1
ADP	var_3	02A	PA	"05'"	"C5'"	-59.942	20.000	1
ADP	var_4	PA	"05'"	"C5'"	"C4'"	179.996	20.000	1
ADP	var_5	"05'"	"C5'"	"C4'"	"C3'"	176.858	20.000	3
ADP	var_6	"C5'"	"C4'"	"04'"	"C1'"	150.000	20.000	1
ADP	var_7	"C5'"	"C4'"	"C3'"	"C2'"	-150.000	20.000	3

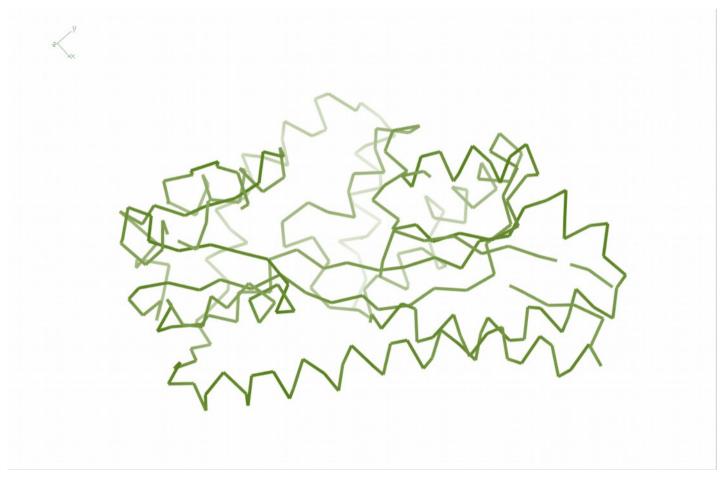
#### Unimodal Torsions and Pseudo Planes



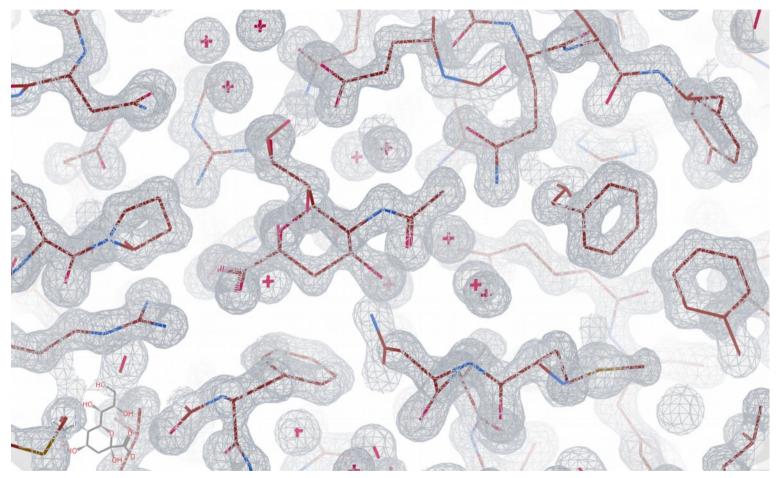


#### External Distance Restraints

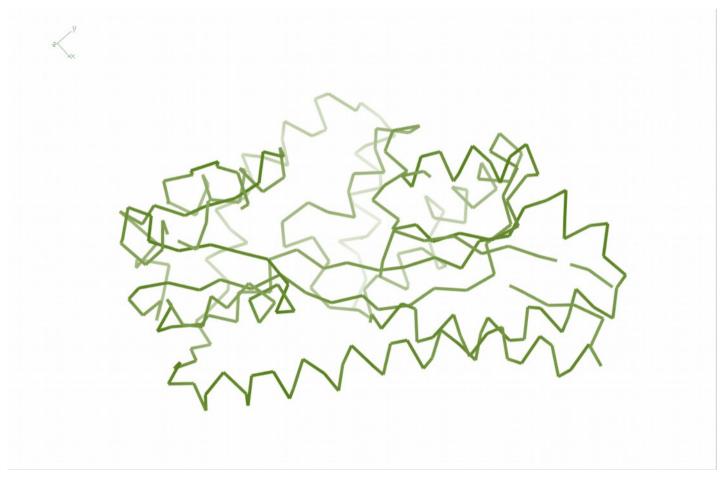
- Using distances from prior-know glycan crystal structures
  - c.f. ProSMART for protein models
  - here we use intent to use a consensus model rather than a particular model



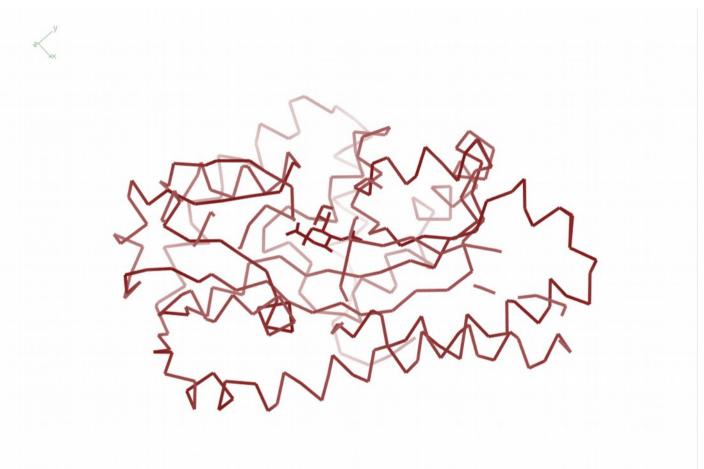
Low resolution current structure



Previously Known High-resolution Reference



Low resolution current structure

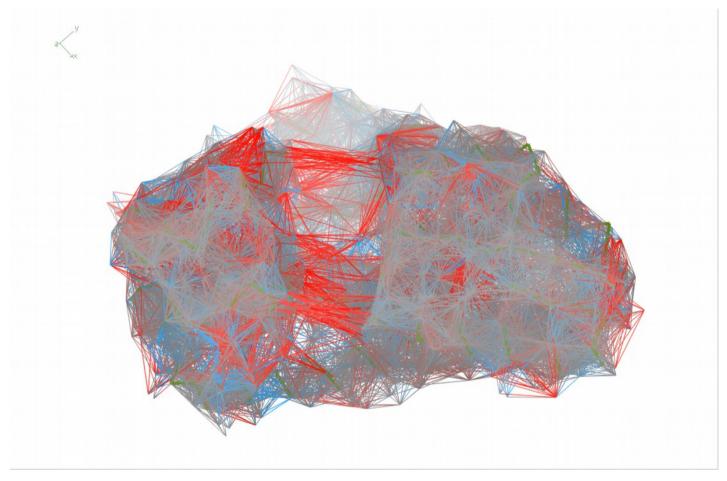


High resolution reference structure

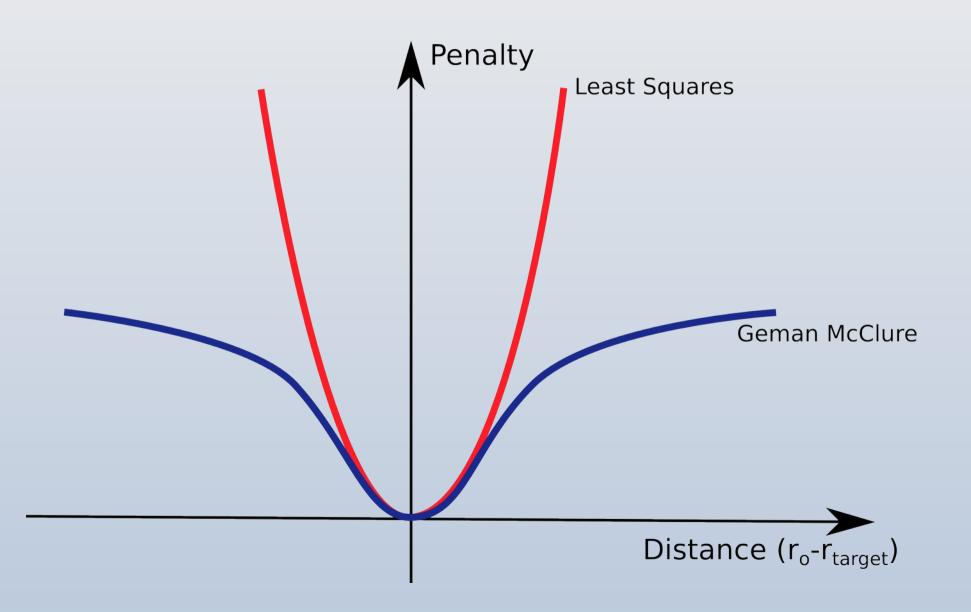
### **ProSMART** Restrains



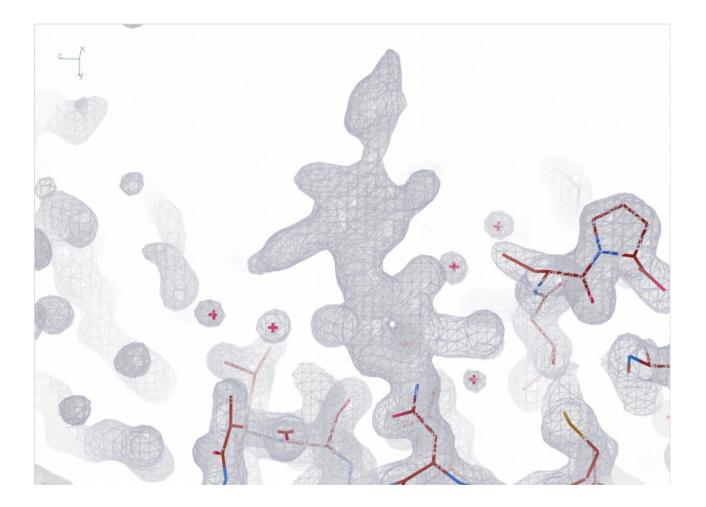
Similar but different



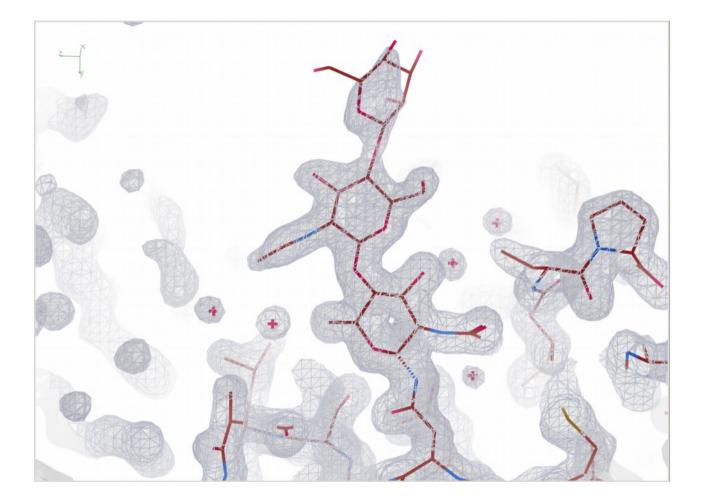
#### **Modified Target Function**



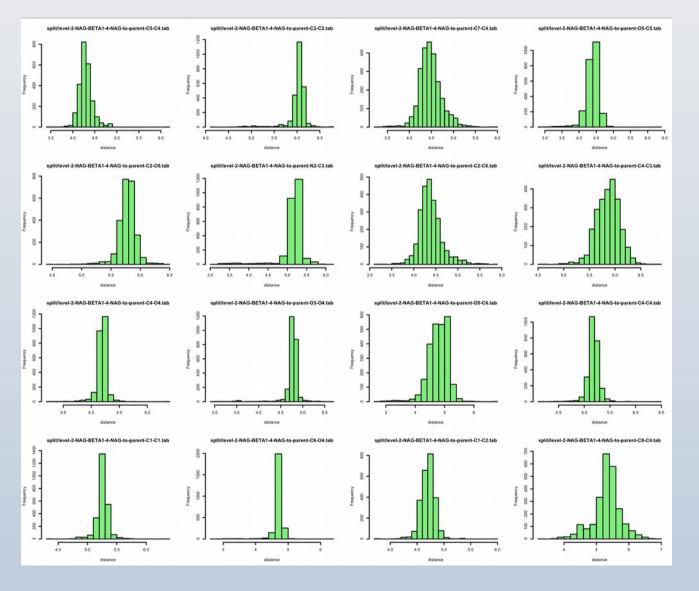
### N-linked Carbohydrate



### N-linked Carbohydrate



### Ligand Tools: N-linked Carbohydrate



 $\rightarrow$  consensus restraints (no user-defined prior)

### Adding PRIVATEER for Model Validation

- Until 2016 *Coot* had no validation for carbohydrate geometry
  - (only fit to density was used)
- Now the model is validated (and filtered) by tree
  - using the output of PRIVATEER
  - both GUI interface and built into the auto-builder
- New Interface

### Video/Demo

### Building Models "Wrongly" (judging by density)

	Good Density	Poor/Bad density
Model built	$\checkmark$	False Positive
No Model	False Negative	$\checkmark$